Progestogens Used in Postmenopausal Hormone Therapy: Differences in Their Pharmacological Properties, Intracellular Actions, and Clinical Effects

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The safety of progestogens as a class has come under increased scrutiny after the publication of data from the Women’s Health Initiative trial, particularly with respect to breast cancer and cardiovascular disease risk, despite the fact that only one progestogen, medroxyprogesterone acetate, was used in this study. Inconsistency in nomenclature has also caused confusion between synthetic progestogens, defined here by the term progestin, and natural progesterone. Although all progestogens by definition have progestational activity, they also have a divergent range of other properties that can translate to very different clinical effects. Endometrial protection is the primary reason for prescribing a progestogen concomitantly with postmenopausal estrogen therapy in women with a uterus, but several progestogens are known to have a range of other potentially beneficial effects, for example on the nervous and cardiovascular systems. Because women remain suspicious of the progestogen component of postmenopausal hormone therapy in the light of the Women’s Health Initiative trial, practitioners should not ignore the potential benefits to their patients of some progestogens by considering them to be a single pharmacological class. There is a lack of understanding of the differences between progestins and progesterone and between individual progestins differing in their effects on the cardiovascular and nervous systems, the breast, and bone. This review elucidates the differences between the substantial number of individual progestogens employed in postmenopausal hormone therapy, including both progestins and progesterone. We conclude that these differences in chemical structure, metabolism, pharmacokinetics, affinity, potency, and efficacy via steroid receptors, intracellular action, and biological and clinical effects confirm the absence of a class effect of progestogens. (Endocrine Reviews 34: 171–208, 2013)

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Progestogens, compounds that exhibit gestational activity, include the only natural progestogen, progesterone, and a variety of synthetic progestogens. In post-
menopausal women, progestogens are used therapeutically for protecting the endometrium against hyperplasia during estrogen therapy. One of the most widely used progestogens for that purpose is medroxyprogesterone acetate (MPA), which has been used for a considerable number of years, either continuously, combined with an estrogen, or sequentially. However, the safety of MPA and that of all other progestogens has been questioned after the results of the estrogen-plus-progestogen and estrogen-alone arms of the Women’s Health Initiative (WHI) trial were published. The data showed increased breast cancer risk with the estrogen/MPA formulation but decreased risk with estrogen alone (1, 2). Although MPA was the only progestogen used in the WHI trial, safety concerns have recently been directed toward progestogens as a general class.

The objective of this review is to determine whether there is any reliable evidence to support the view for a general, uniform effect (class effect) of progestogens. To this end, progestogens will be compared with respect to their chemical structure, structure-function relationships, metabolism, pharmacokinetic parameters, potency, and efficacy via steroid receptors, intracellular mechanism of action, affinity, and biological and clinical effects.

II. Classification of Progestogens

The definition of a progestogen as a compound with progestational activity refers to its action of inducing a secretory endometrium to support gestation. This function of the rising levels of endogenous progesterone after ovulation prepares the endometrium for implantation of a fertilized egg, as well as supporting the uterine lining during a pregnancy, when circulating progesterone reaches characteristically high levels. The term progestogen has been used synonymously with other terms, such as progestagen, gestogen, gestagen, and progestin (3). However, recently, the term progestin has often been used exclusively to describe synthetic progestogens such as MPA, norethindrone, and levonorgestrel, thus excluding the natural progestogen, progesterone. Use of the term progestogen has been consistent with the nomenclature of other hormone groups, such as androgens and estrogens, which are defined as compounds having androgenic and estrogenic activity, respectively. To avoid confusion in light of current practices, the North American Menopause Society has recommended that the term progestogen should be used when referring to progesterone and synthetic progestogens collectively, whereas the name progestin is specific only to synthetic progestogens (4). The nomenclature recommended by North American Menopause Society will be used in the present article.

Progestogens can be divided into two types: natural and synthetic (Table 1) (5). As stated earlier, there is only one natural progestogen, progesterone, which has the chemical structure shown in Fig. 1A. In contrast, there are a variety of progestins that are available for therapeutic use, which vary widely in their chemical structures, as evident in Figs. 2-6. For convenience, these have been classified into two groups: 1) those structurally related to progesterone and 2) those structurally related to testosterone. The chemical structure of testosterone is shown in Fig. 1B. These structural similarities have nothing to do with the actual precursor used to synthesize the progestins, which are derived by multiple chemical reactions from a variety of starting compounds.

Table 1. Classification of progestogens

<table>
<thead>
<tr>
<th>Classification</th>
<th>Progestogen</th>
</tr>
</thead>
<tbody>
<tr>
<td>Natural Progesterone</td>
<td>MPA, megestrol acetate, chlormadinone acetate, cyproterone acetate</td>
</tr>
<tr>
<td>Synthetic</td>
<td>Dydrogesterone, medrogestone</td>
</tr>
<tr>
<td>Structurally related to progesterone</td>
<td>Nomegestrol acetate, nesterone</td>
</tr>
<tr>
<td>19-Norpregnane derivatives</td>
<td>Demegestone, promegestone, trimegestone</td>
</tr>
<tr>
<td>Acetylated</td>
<td></td>
</tr>
<tr>
<td>Nonacetylated</td>
<td></td>
</tr>
<tr>
<td>Acetylated</td>
<td></td>
</tr>
<tr>
<td>Nonacetylated</td>
<td></td>
</tr>
<tr>
<td>Ethinylated</td>
<td>Norethindrone, norethindrone acetate, ethynodiol diacetate, norethynodrel, lynestrenol, tibolone</td>
</tr>
<tr>
<td>Estranes</td>
<td>Levonorgestrel, desogestrel, norgestimate, gestodene</td>
</tr>
<tr>
<td>13-Ethylgonanes</td>
<td>Dinogest, drosiprene</td>
</tr>
<tr>
<td>Nonethinylated</td>
<td></td>
</tr>
</tbody>
</table>

Figure 1.

Figure 1. A, Chemical structure of the natural progestogen, progesterone; B, chemical structure of testosterone.
Progestins structurally related to progesterone can be subdivided into those with and without a methyl group at carbon 10, i.e., pregnane and 19-norpregnane derivatives, respectively. These derivatives are further classified as those that are acetylated and those that are not. Progestins structurally related to testosterone can be subdivided into those that contain an ethinyl group at carbon 17 and those that are nonethinylated. The ethinylated derivatives are further classified as those that have an estrane structure and those that have an 13-ethylgonane structure.

III. Structure-Function Relationships of Progestogens

The biological activity of a progestogen changes considerably, depending on its chemical structure, particularly with respect to pharmacokinetics and potency (5). Structural aspects are discussed below and depicted in Figs. 2–6; differences in pharmacokinetics, potency, and efficacy will be addressed later in this review.

A. Progestogens structurally related to progesterone

1. Pregnan derivatives (Fig. 2)

Starting with progesterone, the addition of a hydroxyl group at carbon 17 renders it devoid of biological progestational activity, but acetylation of that hydroxyl group restores some progestational activity, and the molecule is somewhat active when administered orally. Taking the additional step of adding a methyl group at carbon 6, the resulting molecule, MPA, exhibits relatively high progestational activity and is highly active when given orally (6).

Three highly potent progestogens, megestrol acetate, chlormadinone acetate, and cyproterone acetate, are structurally related to the MPA molecule. Megestrol acetate differs from MPA only in the presence of a double bond between carbons 6 and 7. Chlormadinone acetate and cyproterone acetate have, in addition to a double bond between carbons 6 and 7, a chloral group substituted for the methyl group at carbon 6. Cyproterone acetate differs from chlormadinone acetate only in that it has a methylene group attached to carbons 1 and 2.

Dydrogesterone is one of a group of compounds called retroprogesterones, which have a methyl group at carbon 10 but are not acetylated. They are unique in that the methyl group at carbon 10 is in the α-orientation, instead of the β-orientation seen in progesterone and the pregnanes. Dydrogesterone also has a double bond between carbons 6 and 7, and unlike progesterone, apparently does not inhibit ovulation when given throughout the menstrual cycle and does not alter the basal body temperature (7). These dramatic differences in peripheral and central effects therefore seem to be a consequence of the change in spatial orientation of the methyl group at carbon 10.

Medrogestone, also nonacetylated and having a methyl group at carbon 10, differs from progesterone in that it contains a methyl group at carbons 6 and 17 and a double bond between carbons 6 and 7.

2. 19-Norpregnane derivatives (Fig. 3)

The norpregnane derivatives all lack a methyl group at carbon 10 and include nomegestrol acetate, nesterone, demegestone, promegestone, and trimegestone. Apart from the absence of the methyl group at carbon 10, the norpregnane derivative nomegestrol acetate is identical to that of the pregnane derivative megestrol acetate. Nesterone differs from nomegestrol acetate in the presence of a methylene group at carbon 16 and absence of the methyl group at carbon 6 and the double bond between carbons 6 and 7. Unlike nomegestrol acetate and nesterone, demegestone, promegestone, and trimegestone all have a double bond between carbons 9 and 10 and a methyl group substituted for the acetate group at carbon 17.
Promegestone and trimegestone also have a methyl group on the two-carbon side chain at carbon 17; in trimegestone, the penultimate carbon is also hydroxylated.

### B. Progestogens structurally related to testosterone

#### 1. Ethinylated derivatives: estranes

Starting with the testosterone molecule instead of progesterone, biological activity can be dramatically altered by small changes in the molecular structure. Androgenicity is substantially reduced by the addition of an ethinyl group to form the new compound 17\(\beta\)-ethinyltestosterone, commonly known as ethisterone, which has some progestational and oral activity. These progestogenic and oral activities of ethisterone are further enhanced, and androgenicity almost eliminated, by removal of the methyl group at carbon 10 to form norethindrone (U.S. name), known as norethisterone in Europe and elsewhere.

The norethindrone family of progestogens is referred to as the estranes (Fig. 4) because they all have the same 18-carbon steroid nucleus as the parent steroid, estrane. This family of progestogens also includes norethindrone acetate, ethynodiol diacetate, norethynodrel, and lynestrenol. Norethindrone acetate and ethynodiol diacetate differ from norethindrone by having an acetate group at carbon 3 and at carbons 3 and 17, respectively. Norethynodrel has an identical structure to norethindrone except for a shift in the double bond from carbons 4, 5 of norethindrone to carbons 5, 10 in the norethynodrel molecule. Lynestrenol differs from norethindrone only in the absence of an oxygenated functional group at carbon 3. Tibolone is identical in structure to norethynodrel except it has a methyl group at carbon 7. It is metabolized rapidly and extensively into three metabolites. Two of its metabolites, 3\(\alpha\)- and 3\(\beta\)-hydroxytibolone, bind to the estrogen receptor (ER), whereas the \(\Delta^4\) metabolite binds to the progesterone receptor (PR). Tibolone itself binds with low affinity to the progesterone and androgen receptors.

#### 2. Ethinylated derivatives: 13-ethyl gonanes

When the methyl group at carbon 13 of norethindrone is replaced by an ethyl group, a racemic mixture of \(D(-)-\)norgestrel (levonorgestrel) and \(L(+)-\)norgestrel (dextronorgestrel) results (8). Levonorgestrel is the biologically active form of norgestrel and has proved to be one of the most potent orally active progestogens (7).

The levonorgestrel family of progestogens is sometimes referred to as gonanes; however, this is not appropriate because all steroids by definition are gonanes because they contain the 4-ring carbon nucleus (gonane). A more appropriate name for these progestogens is 13-ethyl gonanes (Fig. 5) (9).

Other progestogens in the levonorgestrel family of 13-ethylgonanes include desogestrel, norgestimate, and gestodene. Having arrived on the scene more recently than levonorgestrel, norethindrone, and progestogens structurally related to norethindrone, these compounds are often referred to as the new progestogens. Desogestrel differs from levonorgestrel by having no oxygenated functional group at carbon 3 but a methylene group at carbon 11, whereas norgestimate has an oxime group at carbon 3 and an acetate group at carbon 17. Gestodene is closer in structure to the parent compound levonorgestrel, merely having an additional double bond between carbons 15 and 16.

#### 3. Nonethinylated derivatives

The nonethinylated subgroup of progestogens consists of the compounds dienogest and drospirenone (Fig. 6). Dienogest is similar in structure to norethindrone except for a cyanomethyl group instead of an ethinyl group at carbon 17 and a double bond between carbons 9 and 10. Drospirenone is structurally related to spironolactone and contains the androstane skeleton to which are attached...
methylene groups at carbons 6 and 7, as well as carbons 15 and 16, and a carbolactone group at carbon 17.

IV. Metabolism of Progestogens

The metabolism of progestogens is poorly understood, largely because relatively few studies on the metabolism of the different progestogens have been carried out.

Progestogens administered orally undergo hepatic first-pass metabolism. The extent to which this occurs varies and depends on the chemical structure of the progestogen. After oral ingestion, progestogens are first subjected to incomplete metabolism by enzymes in intestinal bacteria and the intestinal mucosa. The enzymes include reductases and dehydrogenases, which can add hydrogens to double bonds and ketone groups on progestogen molecules, forming 5α- or 5β-dihydro, 3α- or 3β-hydroxy, and/or 20α- or 20β-hydroxy metabolites.

The metabolized and unmetabolized progestogens are absorbed and enter the portal vein blood at high concentrations. In the liver, they are subjected to a plethora of steroidogenic enzymes, including cytochrome P450 enzymes, which are capable of transforming the metabolized and unmetabolized progestogen molecule into numerous metabolites. Progestogens can also undergo enterohepatic recirculation, but the extent to which this occurs for the different progestogens is poorly understood.

After parenteral administration of a progestogen, the liver is still a major site of progestogen metabolism, even though there is no hepatic first-pass metabolism. The major difference between the metabolism of a drug given orally and one administered parenterally is that the liver is initially exposed to a highly concentrated bolus of unmetabolized and metabolized progestogen.

Of all the studies on metabolism of different progestogens, we know most about progesterone metabolism. Progesterone is highly vulnerable to enzymatic reduction by reductases and hydroxysteroid dehydrogenases during hepatic first-pass metabolism, because its structure contains two ketone groups and a double bond (10). Thus, the molecule is transformed to two isomers of dihydprogesterone, four pregnanolone isomers, and eight isomers of pregnanediol. In addition, progesterone can undergo hydroxylation by cytochrome P450 enzymes. Subsequently, all progesterone metabolites with a hydroxyl group can be sulfated and glucuronidated, and these conjugated products are then excreted in urine and feces. In addition to undergoing extensive transformation during the hepatic first pass, progesterone is also poorly absorbed when administered in a crystalline form. However, when the crystals are broken down to fine particles by the process of micronization, its absorption is improved substantially. The micronization process gives rise to a greater surface area of the compound, allowing it to be dissolved more readily in the aqueous medium of the intestine.

Surprisingly, very little is known about the metabolism of the progestogen most widely used for postmenopausal hormone therapy (HT), i.e. MPA. It has been shown that MPA un-
dergoes ring A reduction, hydroxylation at carbons 6 and 21, and conjugation (primarily glucuronidation) (5). Because ring A of MPA possesses the $\Delta^4$-3-ketone structure found in progesterone, one would expect that the two functional groups would be reduced in a similar manner as those in progesterone; i.e. ring A dihydro and tetrahydro MPA metabolites would be formed. However, unlike progesterone, the reduction of the ketone group at carbon 20 may be impaired due to possible steric hindrance by the acetate group at carbon 17 on the MPA molecule.

Little is also known about the other progestins related in chemical structure to progesterone. However, one would expect those progestins that have a $\Delta^4$-3-ketone structure and/or a ketone group at carbon 20 to undergo reduction in a similar manner as progesterone. Again, reduction may be impaired at carbon 20 in the presence of a functional group (acetate or methyl) at carbon 17 due to steric hindrance.

Relatively more is known about the metabolism of progestins structurally related to testosterone (8). It has been shown that norethindrone and levonorgestrel undergo extensive ring A reactions forming reduced and, to a lesser extent, hydroxylated metabolites. The parent compounds and their metabolites can be conjugated, forming sulfated and glucuronidated products, which are excreted primarily in urine and also in feces. It has also been shown that significant amounts of ethinylestradiol are formed after administration of norethindrone orally to postmenopausal women (11, 12). In fact, it was estimated that oral administration of a 0.5- to 1.0-mg dose of norethindrone combined with ethinylestradiol may add as much as 2–10 $\mu$g ethinylestradiol to the existing dose (11).

What is the biological significance of progestogen metabolites? First, some progestogens are prodrugs and require biochemical transformation to active metabolites. The norethindrone derivatives, which include norethindrone acetate, ethynodiol diacetate, nor ethynodrel, and lynestrenol, have no progestational activity. However, after their oral administration, they are rapidly converted to the progestationally active compound, norethindrone. Desogestrel and norgestimate are also prodrugs. The former compound is converted to the active progestogen etonogestrel (previously called 3-ketodesogestrel), whereas norgestimate is converted to the progestationally active metabolites levonorgestrel and norelgestromin (previously called levonorgestrel-3-oxime). Second, conjugated progestogen metabolites, such as sulfates of norethindrone and levonorgestrel, which are inactive, may form circulating reservoirs from which the active progestogens may be obtained by sulfatase activity. Third, the steroidal milieu consisting of numerous metabolites obtained after administration of a progestogen is unique for each progestogen. Different biological effects may be produced by administered progestogens, due to the specific influence of each progestogen and its metabolites on the conformation of the progestogen receptor and its subsequent activation of transcription in target cells.

V. Pharmacokinetics of Progestogens

Pharmacokinetics (absorption, distribution, and excretion) determine how much of the progestogen administered is available to tissues, primarily by measuring its blood level, and the amount that enters the cells is determined by the extent to which it is bound to carrier proteins that cannot cross the cell membranes. After a progestogen enters the systemic circulation, it is distributed between blood and tissues by passive diffusion. The pattern of distribution of the progestogen is mainly regulated by its binding to transport proteins and tissue receptors. In the blood compartment, all progestogens are bound with low affinity and high capacity to albumin. In addition, some of the progestogens that are structurally related to testosterone also bind with high affinity but low capacity to SHBG; they include norethindrone, levonorgestrel, etonogestrel, and gestodene (13, 14) (Table 2). A relatively smaller amount of progesterone is also bound with high affinity and low capacity, but not to SHBG; instead, it is bound to corticosteroid-binding globulin (15) (Table 2). The binding of progestogens to transport proteins is reversible, so that a change in the concentration of a binding protein in one compartment is followed by a reequilibration of these
compounds in that compartment. Alterations in binding protein concentrations may contribute to the kinetic variability of a progestogen.

It is well recognized that the non-protein-bound (unbound or free) fraction of a steroid is available for metabolism in steroid-metabolizing cells or binding to a receptor in target cells. However, because the binding of steroids to albumin is relatively weak, albumin-bound steroids are also generally considered to be available for metabolism or binding to receptors. There is a paucity of data on free and bioavailable (albumin-bound plus free) fractions of progestogens.

A. Progestogens administered orally

The most common route of progestogen administration for postmenopausal HT and steroidal contraception is oral, yet there is a paucity of information on the pharmacokinetics of progestogens by this route. Progestogens given orally generally reach a maximum concentration within 1–3 h; the maximum concentration and area under the curve depend on the dose. Information on bioavailability and half-life has been derived from frequent blood sampling during 24 h after oral dosing. Bioavailability represents the amount of the progestogen that is found in the circulation after undergoing hepatic first-pass metabolism, estimated by plotting the blood level of the drug against time after administering a given dose both orally and iv and then comparing the areas under the curve; the resulting fraction is multiplied by 100%. Half-life is the time (in hours) over which a drug’s blood level drops to one half of its highest value after dosing. Approximate values taken from the literature (16–31) for bioavailabilities and half-lives of progestogens are summarized in Table 3.

Among progesterone and progestogens structurally related to progesterone, the highest bioavailabilities (>90%) are obtained with MPA, chlormadinone acetate, and trimedestone. In contrast, the bioavailability of progesterone is only less than 5%, and that of dydrogesterone and nomegestrol acetate is 28 and 60%, respectively. Chlormadinone acetate, cyproterone acetate, and nomegestrol acetate have the longest half-lives (80.1, 54.0–78.6, and 50 h, respectively), whereas that of medrogestone is substantially lower (34.9 h). Progesterone and other progestogens related to progesterone (including MPA, megestrol acetate, dydrogesterone, and trimedestone) have even shorter half-lives, ranging from 15–24 h.

Among progestogens structurally related to testosterone, the highest bioavailabilities are achieved with levonorgestrel, gestodene, and dienogest, reaching more than 90%, whereas norethindrone, desogestrel, and drospirenone have bioavailabilities in the range of 62–76%. The longest half-life occurs with drospirenone (31.1–32.5 h), whereas norethindrone has the shortest (8 h); interme-

### Table 2. Distribution of progestogens bound to SHBG or CBG in blood

<table>
<thead>
<tr>
<th>Progestogen (Ref.)</th>
<th>SHBG-bound (%)</th>
<th>CBG-bound (%)</th>
<th>Albumin-bound (%)</th>
<th>Free (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Norethindrone (13)</td>
<td>35.5</td>
<td>ND</td>
<td>60.8</td>
<td>3.7</td>
</tr>
<tr>
<td>Levonorgestrel (13)</td>
<td>47.5</td>
<td>ND</td>
<td>50.0</td>
<td>2.5</td>
</tr>
<tr>
<td>Etonogestrel (14)</td>
<td>31.6</td>
<td>ND</td>
<td>65.9</td>
<td>2.5</td>
</tr>
<tr>
<td>Gestodene (14)</td>
<td>75.3</td>
<td>ND</td>
<td>24.1</td>
<td>0.6</td>
</tr>
<tr>
<td>Progesterone (15)</td>
<td>0.6</td>
<td>17.7</td>
<td>79.3</td>
<td>2.4</td>
</tr>
</tbody>
</table>

CBG, Corticosteroid-binding globulin; ND, not detected.

### Table 3. Average bioavailabilities and half-lives of progestogens

<table>
<thead>
<tr>
<th>Progestogen</th>
<th>Dose (mg)</th>
<th>Bioavailability (%)</th>
<th>Half-life (h)</th>
<th>Ref.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Progesterone</td>
<td>100, 200, 300</td>
<td>&lt;5</td>
<td>16.2–18.3</td>
<td>16</td>
</tr>
<tr>
<td>MPA</td>
<td>10</td>
<td>&gt;90</td>
<td>24</td>
<td>17</td>
</tr>
<tr>
<td>Megestrol acetate</td>
<td>160</td>
<td>NA</td>
<td>22.3</td>
<td>18</td>
</tr>
<tr>
<td>Cyproterone acetate</td>
<td>2</td>
<td>NA</td>
<td>54.0–78.6</td>
<td>19</td>
</tr>
<tr>
<td>Chlormadinone acetate</td>
<td>2</td>
<td>~100</td>
<td>80.1</td>
<td>20</td>
</tr>
<tr>
<td>Medrogestone</td>
<td>5</td>
<td>NA</td>
<td>34.9</td>
<td>21</td>
</tr>
<tr>
<td>Dydrogesterone</td>
<td>10</td>
<td>28</td>
<td>14–17</td>
<td>—a</td>
</tr>
<tr>
<td>Nomegestrol acetate</td>
<td>2.5</td>
<td>60</td>
<td>50</td>
<td>22</td>
</tr>
<tr>
<td>Trimegestone</td>
<td>0.5</td>
<td>~100</td>
<td>15</td>
<td>—a</td>
</tr>
<tr>
<td>Norethindrone</td>
<td>1</td>
<td>64</td>
<td>8</td>
<td>23</td>
</tr>
<tr>
<td>Levonorgestrel</td>
<td>0.15–0.25</td>
<td>89/99b</td>
<td>9.9/13.2b</td>
<td>24</td>
</tr>
<tr>
<td>Desogestrel</td>
<td>0.15</td>
<td>62/76b</td>
<td>11.9/23.8b</td>
<td>25, 26</td>
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<tr>
<td>Gestodene</td>
<td>0.075</td>
<td>87/99b</td>
<td>12–14</td>
<td>27, 28</td>
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<tr>
<td>Dienogest</td>
<td>4</td>
<td>96.2</td>
<td>10.8/11.6b</td>
<td>29</td>
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<tr>
<td>Drospirenone</td>
<td>3</td>
<td>66</td>
<td>31.1–32.5</td>
<td>30, 31</td>
</tr>
</tbody>
</table>

NA, No data available.

a The data were obtained from a package insert.

b Multiple bioavailability or half-life values are shown.
Concentrations in biopsies taken after treatment of women with progesterone creams were higher than those in biopsies from women treated with oral progesterone. In the vaginal administration group, serum progesterone levels rose more rapidly, plateauing at about 16 ng/ml after 4 h of treatment. In the im group, serum progesterone levels rose more slowly and reached a peak of about 7 ng/ml after 4 h. Circulating progesterone levels in biopsies taken after 7 d of treatment were considerably higher after intravaginal than im dosing, despite the higher serum progesterone levels after im injection. This study highlights the potential importance of the vaginal route in menopausal HT, because the endometrium is the most important target of progesterone action in this application.

3. Percutaneous route

The use of progesterone in the form of transdermal delivery via topical creams or gels has been a subject of some concern because of speculation that the low serum progesterone levels achieved with these agents indicate an insufficient secretory effect on the endometrium (34). However, despite such low serum levels below 4 ng/ml, antiproliferative effects on the endometrium have been demonstrated with progesterone creams (35), and in addition, salivary progesterone levels are found to be very high (36), indicating that progesterone levels in serum do not necessarily reflect those in tissues. The effects of topical progesterone creams on the endometrium should therefore be based on histological examination of the endometrium rather than on serum levels.

An important caveat with progesterone cream products that are readily available over the counter is that some of these products do not contain progesterone but instead contain wild yam extract in which the precursor for the synthesis of progesterone, diosgenin, is present. However, the chemical reactions required to convert the diosgenin in wild yam extract to progesterone can be carried out only in a laboratory and do not occur in the body.

Two different progestins, levonorgestril and norethindrone acetate, are used in different transdermal systems, each in combination with estradiol. Both systems are adhesive-based matrix transdermal patches designed to release estradiol and levonorgestril or norethindrone acetate continuously for 7 or 3.5 d, respectively. The levonorgestril/estradiol-containing system (Climara Pro) provides a levonorgestril nominal delivery rate of 0.015 mg/d (37). After its application, in one study, levonorgestril concentrations were maximal after approximately 2.5 d, and average serum steady-state concentrations were 166 pg/ml (38). The norethindrone acetate/estradiol-containing system (CombiPatch) is available in two different doses of the progestin, with nominal delivery rates of 0.14 and 0.25 mg/d. In one study, norethindrone steady-state concentrations were attained within 24 h of application and the subsequent average serum steady-state concentrations were 489 and 840 pg/ml for the respective doses (39).

C. Drug interactions

The potential interaction of progestogens with other drugs has been the subject of numerous reports since the early 1970s. Some interactions are well documented and therapeutically relevant; however, many remain unproven or are the subject of continuing controversy. Strong evidence indicates that griseofulvin (an antifungal drug), rifampin (an antituberculosis drug), and certain anticonvulsants (phenobarbital and phenytoin) induce hepatic enzymes and decrease oral contraceptive (OC) effective-
ness. An unproven, but widely accepted, drug interaction involves the effect of antibiotics on OC efficacy. Despite a number of reports implicating penicillins, tetracyclines, and other antibiotics in causing OC failure, no firm evidence links antibiotic administration with altered circulating levels of progestogens.

VI. Intracellular Mechanisms of Action of Progestogens

A. Steroid receptor structure, distribution, and ligand binding

The intracellular actions of progestogens are mediated predominantly via the PR, a ligand-activated transcription factor and member of the steroid receptor and nuclear receptor families of receptors (40). Progestins are designed to be potent, high-affinity PR agonists that mimic the actions of progesterone but with better bioavailability. However, many progestins bind to other members of the steroid receptor family, which includes the androgen receptor (AR), glucocorticoid receptor (GR), and mineralocorticoid receptor (MR), and exhibit off-target effects via these receptors (41, 42). Progestogens do not bind to the ER, the other member of the steroid receptor family. Moreover, current progestogens exhibit considerable variation in their binding affinities via the AR, MR, and GR.

It is not surprising that progestogens cross-react with several members of the steroid receptor family, because the PR, AR, GR, MR, and ER share significant amino acid homology in certain regions, while exhibiting a highly conserved overall domain structure. These domains include an unconserved amino-terminal domain of variable length, a highly variable transcriptional activation function-1 (TAF-1) domain situated near the N terminus, a highly conserved DNA-binding domain (DBD), as well as a moderately conserved C-terminal ligand-binding domain (LBD). The TAF-1 domain has been reported to be ligand independent and required for optimal transcriptional activity via protein-protein interactions with general transcription factors as well as cofactors (43). The DBD, the most conserved domain of the steroid receptors, contains two zinc finger motifs and is responsible for sequence-specific and high-affinity DNA binding, as well as playing a role in receptor dimerization, interaction with cofactors (44), and nuclear localization (45). The LBD, toward the C terminus, determines ligand specificity and affinity, as well as playing a role in dimerization, nuclear localization, and interaction with chaperone proteins and cofactors (45–47). A highly conserved TAF-2 domain is present within the LBD, which contains at least one co-factor interaction motif important for ligand-dependent transcriptional activity (46, 48). Despite the approximately 50–60% amino acid sequence homology between the LBDs of the PR, AR, GR, and MR, these steroid receptors exhibit subtle differences in their dimerization and cofactor binding sites due to differences in secondary structure, whereas the ER is even less conserved (49–54).

Progestogen action via steroid receptors is further complicated by the presence of several receptor isoforms for each receptor. The PR exists as two isoforms, PR-A and PR-B, transcribed from two promoters of a single gene (55). The longer PR-B isoform is more transcriptionally active and contains a third transactivation function domain that is absent from PR-A, allowing binding of co-activators to PR-B that do not bind to PR-A (56–58). Similarly, other steroid receptors exist in several isoforms that exhibit differential expression profiles and functions (40, 59, 60). The PR, ER, AR, and MR have a relatively selective distribution. The PR is expressed in the female reproductive tract, mammary gland, brain, and pituitary gland as well as some immune-function cells (61, 62). Ratios of the individual PR isoforms vary in the ovary, breast, and uterus (63), where they have different physiological functions in various target cells (63, 64), most likely due to the distinct and promoter-specific transactivation effects of PR-A and PR-B (65). Changes in the ratio of PR-A to PR-B have been implicated in the development of breast cancers, most likely via a mechanism involving MAPK-dependent PR phosphorylation and isoform stability (66). Changes in PR isoform expression levels and/or activity have also been associated with functional progesterone withdrawal in the human pregnant uterus (67). The two main ER isoforms, ERα and ERβ, have distinct tissue expression patterns and roles in disease and normal physiology in breast, ovary, colon, endometrium, and bone cells in women (68). The AR is expressed in the mammary gland, muscle, prostate, skin, vagina, bone marrow, and testes (40). Thus, AR effects are likely to be responsible for differential progestogen actions in these tissues, particularly in the breast. In contrast, the GR is ubiquitously expressed, although its levels are regulated in a tissue- and cell-cycle-specific manner (40). Therefore, differential progestogen effects mediated by the GR are likely to occur in most tissues and in particular those where GR levels are high, such as in immune-function cells. Interestingly, GR levels have been shown to vary widely in different breast carcinoma subtypes (69), suggesting a particularly important role of varying GR levels in the determination of effects of progestogens such as MPA in breast cancer. Differential expression profiles and functions of GR isoforms, such as GRα and GRβ (59), would increase the possibilities for differential progestogen actions via the GR.
MR, although not as widely expressed as the GR, is also expressed in many tissues, including the kidney, colon, central nervous system, heart, adipocytes, and vascular cells (40, 70–72). Thus, physiological functions in these tissues are likely to be modulated selectively by progestogens via the MR.

To determine the affinity of a progestogen for a particular receptor, binding studies have been developed. These have been performed in a wide range of models including animal or human tissue, human cell lines expressing endogenous receptors, cell lines deficient in endogenous receptors but overexpressing exogenous human steroid receptors, or even in \textit{in vitro} systems using recombinant purified human receptor.

Binding assays are usually performed using a constant concentration of radiolabeled reference agonist incubated with varying concentrations of unlabeled competitor ligand to obtain an IC\textsubscript{50} for the competitor steroid. Affinities are usually expressed as relative binding affinity (RBA), which is calculated by dividing the IC\textsubscript{50} of the test steroid by the IC\textsubscript{50} of the reference steroid, multiplying by 100, and expressing the RBA as a percentage. The IC\textsubscript{50} is the concentration of the unlabeled steroid that corresponds to 50% inhibition of the total binding of the radiolabeled reference agonist. RBAs are often only an approximate measure of relative affinity because IC\textsubscript{50} can vary with receptor concentration, concentration of radiolabeled steroid, and whether or not equilibrium has been reached for both steroids. More accurate affinities can be obtained by determination of time to reach equilibrium for the steroids under investigation as well as by performing homologous and heterologous displacement assays with determination of equilibrium dissociation constants using the Cheng-Prusoff equation or by saturation binding analysis (41).

From Table 4, which summarizes some of the available data on RBAs of progestogens to different steroid receptors, it is immediately apparent that the data show a wide variability. One of the reasons for this is undoubtedly due to different methods used to determine affinity, as discussed above. Another source of variability is the use of different cell or tissue models, which vary in the relative concentrations of different steroid receptors. Off-target binding of the progestogen to receptors other than the one under investigation could effectively lower the apparent RBA, especially if the progestogen has a relatively high affinity for a competing receptor, because the concentration of unlabeled competitor progestogen available for binding to the target receptor will be effectively less than the added concentration. Thus, experiments that determine equilibrium dissociation constants and those using cell lines deficient in endogenous receptors and overexpressing exogenous human steroid receptors or even \textit{in vitro} systems using recombinant purified human receptor are likely to yield more accurate results. Another source of variability is the species from which tissue is obtained as well as the variation in age and pretreatment of the animal or human donor. Note that direct comparisons between the values determined by competition binding using different reference radiolabeled agonists \textit{e.g.} progesterone \textit{vs.} promegestone for the PR or dihydrotestosterone (DHT) \textit{vs.} mibolerone for the AR) for a particular receptor and competitor ligand cannot be made. Nevertheless, despite these sources of error and variability in binding experiments, several valuable insights have been obtained.

Although all progestogens bind with high relative affinity to the PR, most bind with a greater affinity than progesterone (Table 4). As the natural progestational agent of all mammals, progesterone was an obvious choice as the reference steroid for many binding assays and was used in conjunction with [\textsuperscript{3}H]progesterone in competitive binding studies with PRs. More recently, the highly potent synthetic progestin, promegestone (R5020), has replaced progesterone as the reference compound because most progestins have greater progestational activity than progesterone itself. Human and animal tissues can show profound differences in RBAs for the PR. RBAs for norgestimate and its principal active metabolites for uterine PRs were determined in two studies (75, 76); in one (75), norgestimate was bound to the PR in rabbit uterine tissue with an RBA of 124%, whereas norelgestromin and levonorgestrel had RBAs of 94 and 541%, respectively. In the other study (76), which used human uterine tissue instead of rabbit, norgestimate showed very little binding to the PR (RBA, 0.8%) and the binding of norelgestromin was low (RBA, 8%), whereas the RBA of levonorgestrel was 250%. This illustrates the difficulties of extrapolating animal RBA data to human tissues.

Progestogens vary greatly in their reported affinities for the AR, with some of the older-generation progestins such as MPA, norethindrone, and levonorgestrel binding with high affinity relative to testosterone (77–86), although some researchers report similar affinities for progesterone, MPA, norethindrone acetate, and DHT for the AR (Table 4). In contrast, drospirenone, dienogest, and trimegestone exhibit low RBA (74, 87, 88), although reported relative values differ for several progestogens, whereas nesterone does not bind at all to the AR (89).

Progesterone, trimegestone, and drospirenone have a relatively high affinity for the MR (Table 4) (90–93). The latter two progestogens were developed for their antimineralocorticoid properties for contraceptive usage (94) and for their predicted beneficial effects on blood pressure and cardiovascular function (31, 90, 95, 96). However, other progestins such as MPA and norethindrone acetate bind
TABLE 4. RBAs and hormonal activities of progestogens via the PR, AR, GR, and MR

<table>
<thead>
<tr>
<th>Progestogen</th>
<th>PR RBA (%)</th>
<th>Androgenic activity</th>
<th>Antiandrogenic activity</th>
<th>AR RBA (%)</th>
<th>Androgenic activity</th>
<th>Antiandrogenic activity</th>
<th>GR RBA (%)</th>
<th>Glucocorticoid activity</th>
<th>MR RBA (%)</th>
<th>Antimineralocorticoid activity</th>
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<tr>
<td>Progesterone</td>
<td>50</td>
<td>0</td>
<td>(+)</td>
<td>10</td>
<td>0</td>
<td>(+)</td>
<td>100</td>
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<td>+</td>
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<td>100</td>
<td>3</td>
<td>0</td>
<td>(+)</td>
<td>11</td>
<td>1</td>
<td>(+)</td>
<td>100</td>
<td>0</td>
<td>100</td>
<td>+</td>
</tr>
<tr>
<td>Chloromadinone acetate</td>
<td>67</td>
<td>5</td>
<td>+</td>
<td>8</td>
<td>0</td>
<td>+</td>
<td>9</td>
<td>—</td>
<td>9</td>
<td>—</td>
</tr>
<tr>
<td>Cyproterone acetate</td>
<td>90</td>
<td>6</td>
<td>+</td>
<td>6</td>
<td>0</td>
<td>+</td>
<td>8</td>
<td>—</td>
<td>8</td>
<td>—</td>
</tr>
<tr>
<td>Dienogest</td>
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<td>10</td>
<td>—</td>
<td>1</td>
<td>0</td>
<td>—</td>
<td>0</td>
<td>+</td>
<td>0</td>
<td>—</td>
</tr>
<tr>
<td>Drosiprenone</td>
<td>35</td>
<td>5</td>
<td>+</td>
<td>6</td>
<td>0</td>
<td>+</td>
<td>230</td>
<td>+</td>
<td>230</td>
<td>+</td>
</tr>
<tr>
<td>19</td>
<td>2</td>
<td>1</td>
<td>+</td>
<td>0</td>
<td>0</td>
<td>—</td>
<td>500</td>
<td>+</td>
<td>500</td>
<td>+</td>
</tr>
<tr>
<td>Gestodene</td>
<td>90</td>
<td>85</td>
<td>—</td>
<td>27</td>
<td>0</td>
<td>—</td>
<td>290</td>
<td>—</td>
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<tr>
<td>864</td>
<td>71</td>
<td>38</td>
<td>58</td>
<td>1</td>
<td>75</td>
<td>—</td>
<td>380</td>
<td>100</td>
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<tr>
<td>Levonorgestrel</td>
<td>150</td>
<td>45</td>
<td>+</td>
<td>1</td>
<td>7.5</td>
<td>+</td>
<td>175</td>
<td>+</td>
<td>175</td>
<td>+</td>
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<tr>
<td>MPA</td>
<td>115</td>
<td>5</td>
<td>+</td>
<td>29</td>
<td>160</td>
<td>—</td>
<td>31</td>
<td>0.08</td>
<td>31</td>
<td>0.08</td>
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<tr>
<td>Nestorone</td>
<td>136</td>
<td>0</td>
<td>—</td>
<td>38</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
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<tr>
<td>Nomegestrol acetate</td>
<td>125</td>
<td>42</td>
<td>+</td>
<td>6</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
</tr>
<tr>
<td>Norethindrone</td>
<td>75</td>
<td>15</td>
<td>+</td>
<td>0</td>
<td>0</td>
<td>—</td>
<td>0</td>
<td>—</td>
<td>0</td>
<td>—</td>
</tr>
<tr>
<td>134</td>
<td>55</td>
<td>1.4</td>
<td>—</td>
<td>2.7</td>
<td>0.07</td>
<td>—</td>
<td>0.07</td>
<td>—</td>
<td>0.07</td>
<td>—</td>
</tr>
<tr>
<td>Norethindrone acetate</td>
<td>ND</td>
<td>1.7</td>
<td>—</td>
<td>1.6</td>
<td>0.07</td>
<td>—</td>
<td>0.07</td>
<td>—</td>
<td>0.07</td>
<td>—</td>
</tr>
<tr>
<td>Promegestone</td>
<td>100</td>
<td>0</td>
<td>—</td>
<td>5</td>
<td>53</td>
<td>—</td>
<td>53</td>
<td>—</td>
<td>53</td>
<td>—</td>
</tr>
<tr>
<td>Trimegestone</td>
<td>330</td>
<td>1</td>
<td>—</td>
<td>9</td>
<td>120</td>
<td>—</td>
<td>120</td>
<td>—</td>
<td>120</td>
<td>—</td>
</tr>
<tr>
<td>588</td>
<td>2.4</td>
<td>13</td>
<td>—</td>
<td>42</td>
<td>42</td>
<td>—</td>
<td>42</td>
<td>—</td>
<td>42</td>
<td>—</td>
</tr>
</tbody>
</table>

RBAs were determined by competitive binding assays using a radiolabeled reference ligand and increasing concentrations of unlabeled competitor ligand and are based on IC50 values in most cases (a and b), whereas IC50 (equilibrium dissociation constant for an unlabeled competitor or inhibitor ligand competing for binding of the radiolabeled reference ligand to the receptor) values were determined by homologous and heterologous displacement and using the Cheng-Prusoff equation (c) (73). There is no evidence of significant direct binding for any of these steroids to the ER (RBAs all 0 to <1% relative to estradiol) (7, 41). Hormonal activities are based on animal experiments and taken from Refs. 7 and 41. All the steroids are progestogenic, and all exhibit antitestogenic activity in animal models via a mechanism independent of the progestin binding to ER. None of them, except norethindrone, exhibits estrogenic activity (7). Key to activity levels: +, effective; ( +), weakly effective; —, not effective; ? literature inconsistent. ND, Not determined.

A Values were compiled by cross-comparisons from several competitive binding studies that used different methods and were taken from Ref. 7. Most of the data are from animal tissues or cell lines expressing several receptors, and hence, some are likely to be inaccurate. The reference radiolabeled ligands (100% RBA) were as follows: PR, promegestone; AR, metribolone or R1881; GR, dexamethasone; MR, aldosterone.

b Values were determined using recombinant human receptor binding in vitro (74). The reference radiolabeled ligands (100% RBA) were as follows: PR, progesterone; AR, testosterone; GR, dexamethasone; MR, aldosterone.

c RBAs were calculated from Kᵢ (equilibrium dissociation constant for an unlabelled competitor or inhibitor ligand competing for binding of the radiolabeled reference ligand to the receptor) values, determined by expressing the human recombinant GR in the A549 cell line (73) or the human recombinant AR or MR (314) in the COS-1 cell line, both deficient in steroid receptors, using the methods outlined in Ref. 73. The reference ligands (100% RBA) were as follows: AR, mibolerone; GR, dexamethasone; MR, aldosterone. Note that for the AR, the RBA for DHT in this assay was 1.3%.

Weakly to the MR (41), whereas several progestins such as dinogest, nomegestrol acetate, and promegestone do not bind at all (87, 97).

In contrast to PR and AR binding, relatively few progestogens bind to the GR with affinities in the significant pharmacological range, with the notable exceptions of MPA, gestodene, and nestorone (Table 4). MPA has a high RBA for the GR (73, 77, 81, 98–100), and it has been shown that MPA displays significantly higher binding affinity toward the GR than cortisol, the endogenous glucocorticoid in humans (100). Gestodene binds with a relatively high affinity to the GR (101). However, progestins such as norethindrone, levonorgestrel, dinogest, and trimegestone, like progesterone, bind the GR with low relative affinity (31, 73, 74, 82, 87, 88, 99, 100).

In summary, a major determinant of differential intracellular progestogen actions is the variable affinity of progestogens for binding to the PR and to other members of the steroid receptor family. Although all progestogens bind with relatively high affinity to the PR, they do not bind to the ER, and their reported relative affinities for the AR, GR, and MR differ substantially. Affinities, together with concentrations of progestogens and competing endogenous ligands, determine receptor occupancy for a particular steroid receptor. Fractional occupancy is in turn a major determinant of the biological response. Although the equilibrium dissociation constants for a particular progestogen or endogenous ligand for a particular steroid receptor do not change (41), the fractional occupancy of a receptor changes depending on ligand concentration,
which in turn varies according to its relative affinity for, and concentrations of, the different steroid receptors present. Although useful binding data are available, much of it may be inaccurate; additional experiments are required to more accurately determine equilibrium binding constants for most of the progestogens for different steroid receptors and their isoforms, in the absence of confounding factors such as the sources of the receptors, the methods of binding analysis, and the presence of off-target receptors. Given that the relative levels of different receptors and their isoforms vary greatly in different tissues, this is also likely to be a major determinant of differential actions via progestogens.

B. Potency, efficacy, and biocharacter of progestogens via steroid receptors

Progestogens exhibit considerable variation in their potencies and efficacies as well as the resulting extent of agonist, partial agonist, or antagonist responses, i.e., their biocharacter, via steroid receptors. Potency is defined in this context as the concentration of ligand required for half of the maximal biological response, whereas efficacy is the maximal induced response for that particular ligand (41). Agonists, partial agonists, and antagonists all bind to a particular receptor, with an agonist resulting in an efficacy similar to that of the natural ligand, whereas a partial agonist gives a similar response to that of the natural ligand but with a lower efficacy, and an antagonist inhibits the response of an agonist. Partial agonists and antagonists can exhibit varying degrees of antagonism depending on the relative concentrations of competing ligands and their affinities for a particular receptor as well as on receptor concentration.

Much of the data on potency, efficacy, and biocharacter via different steroid receptors has been obtained from animal experiments (41) (Table 4). These data do reflect to some extent the actions of a progestogen via a particular steroid receptor but also suffer from the same source of variability as the binding studies when it comes to off-target effects via other receptors, which would lead to inaccurate potency estimates. In addition, the animal data are also confounded by pharmacokinetic factors, metabolism, binding to serum proteins, and indirect actions of the progestogens via target proteins other than steroid receptors.

Bioassays have been developed that test the effects of progestogens on uterine glandular proliferation, pregnancy maintenance, delay of parturition, or inhibition of ovulation in rabbits or rats. The Clauberg test is based on initial observations made by Clauberg in the 1920s and is the most widely used bioassay for progestational agents. It was later developed into specific protocols by McPhail in 1934 (102). The principle of the test is to measure glandular proliferation in rabbit endometrium that has been primed with estrogen, in response to progestogens given orally or parenterally. McPhail used a standardized scale for grading the complex glandular proliferation of the rabbit endometrium in response to the different progestogens. This scale starts from 0, corresponding to no glandular development, with a highest possible value of +4, corresponding to maximal glandular development. In practice, progestogens are compared at a dose level that produces a value of +2 on the McPhail scale.

The Clauberg test is, however, subject to considerable variation in estimates of potency (103). Problems arise in interpretation of the test because dose-response curves for commonly employed test substances are not parallel. Other commonly used bioassays also have various limitations (103). For example, bioassays that measure pregnancy maintenance as a progestational effect cannot use estrogens, which will inhibit the active progestogens when given at sufficient doses; the bioassay for delay of parturition cannot distinguish between the various progestogens; and the ovulation inhibition bioassay in the laboratory gives different progestogen potencies when compared with those obtained in women. Despite these limitations, bioassays have led to significant insights into progestogen actions, although they frequently do not correlate with the steroid receptor-binding affinity data, in particular for the AR, GR, and MR.

Several general trends have emerged from both the animal bioassays and in vitro binding affinity studies. Although all progestogens bind to the PR (Table 4) and act as progestosterone agonists, they exhibit differences in the potency of the progestogenic responses (Table 4) (104–108). On the other hand, progestogens exhibit a wide spectrum of activities via the AR, ranging from no effect to agonist, partial agonist, and antiandrogenic activity (Table 4). For example, some of the older-generation progestins such as MPA, norethindrone acetate, norethindrone, and levonorgestrel, which bind with relatively high affinity to the AR, have been reported to act as agonists or partial agonists in some contexts, unlike progesterone (Table 4) (77–86), although the androgenic biological activities reported for MPA and progesterone vary greatly in the literature. In contrast, drospirenone, dienogest, and trimgestone, which exhibit low RBA for the AR, exhibit no AR-mediated agonist activity but exhibit variable to potent antiandrogenic properties (Table 4) (74, 87, 88). Nestorone has no activity via the AR (Table 4) (89), whereas nomegestrol acetate, which binds the AR, has no agonist activity and displays partial antiandrogenic activity (Table 4) (22, 109, 110). Consistent with their binding activities, MPA has partial to full agonist activity via the GR in some contexts (41), whereas gestodene ex-
hibits partial agonist activity in some contexts (41). However, progestogens such as norethindrone, levonorgestrel, dienogest, and trimegestone show no or very little glucocorticoid-like activity in most contexts, whereas the reported effects of progesterone via the GR vary (Table 4) (41). Certain progestogens like trimegestone and drospirenone with a relatively high affinity for the MR exhibit weak partial MR agonist activity. However, both progesterone and drospirenone exhibit potent antagonist activity toward aldosterone via the MR, whereas the reported antagonistic effects of trimegestone are variable (Table 4) (90–93). Other progestins such as MPA and norethindrone acetate, which bind weakly to the MR, exhibit no antimineralocorticoid activity in rat models (Table 4) (41), whereas dienogest neither binds to nor displays agonist or antagonist activity for the MR (Table 4) (87, 97).

In addition to in vitro binding affinity tests and bioassays, clinical tests have been used to assess the relative biological effects of progestogens in women; they include those based on delay of menses, induction of secretory changes in the endometrium, inhibition of ovulation, and changes in vaginal cytology and cervical mucus. Traditionally, in these clinical tests, the term potency is often used to refer to a relative response obtained at a chosen progestogen dose, using equivalent mass doses, without dose-response analysis. Alternatively, some assays refer to potency as the comparative dose (in mass) required to give a particular level of response, usually not a maximal response. Hence, these are not true measures of potency or efficacy in terms of the definitions discussed above. Thus, the term potency reported from such clinical studies needs to be interpreted with these limitations in mind. Greenblatt and co-workers (111) were the first to describe the delay-of-menses test for progestogenic potency. In this test, the progestogen is administered beginning on the sixth or seventh day after ovulation and continuing for 3 wk or more. If the progestogen is effective, it will delay menstrual bleeding until 2–3 d after treatment is discontinued. The delay-of-menses test was further developed and standardized by Swyer and Little (112) for assessing comparative potency of progestogens and is consequently referred to as the Swyer-Greenblatt test.

A literature review published in 1985 assessed the relative potency of progestogens used in oral contraception in the United States on the basis of available human data showing the effect of progestogens on the delay of menses by the Swyer-Greenblatt test as well as effects on subnuclear vacuolization (as an indirect determination of glycogen deposition) and lipid and lipoprotein levels (113). The review concluded that norethindrone, norethindrone acetate, and ethynodiol diacetate are approximately equivalent in potency, whereas norgestrel and its bio-

logically active enantiomer, levonorgestrel, are about 5–10 and 10–20 times as potent as a similar weight of norethindrone, respectively. However, there are limitations in the studies that were reviewed. Parallelism of dose-response curves was not demonstrated in the delay-of-menses test, and high doses of ethinyl estradiol (50 and 100 µg) were used in this test and in the subnuclear vacuolization test. Also, only relative effects were obtained in the lipid/lipoprotein tests because the results were not obtained from dose-response curves.

In another approach to determine progestogen potency from clinical data, a series of studies by King and co-workers (114–120) assessed progestogenic effects by analyzing biochemical and morphological features of endometrium from estrogen-primed postmenopausal women. First, the postmenopausal women were treated daily with either 0.625 or 1.25 mg of conjugated equine estrogens, and then the effects of 6 d of sequential progestogen treatment during the last 6–12 d of the month were assessed. At least three different doses of each of 5 orally administered progestogens, specifically, norethindrone, levonorgestrel, MPA, dydrogesterone, and progesterone, were studied. The endometria were analyzed for biochemical parameters including nuclear estradiol receptor levels, DNA synthesis, and isocitric and estradiol dehydrogenase activities. King and Whitehead (121) re-examined the results of these studies to allow comparisons with corresponding premenopausal secretory-phase values and reported the potency of progestogens relative to a value of 1 for norethindrone. The analysis showed that the potency of levonorgestrel was 8-fold greater, whereas the potencies of MPA, dydrogesterone, and progesterone were 10, 50, and 500 times lower, respectively.

The recommended oral progestogen doses for endometrial protection (Table 5) are based on the potencies established by the analysis of King and Whitehead (121); they are 1, 0.15–0.5, 2.5–10, 20, and 100–300 mg for norethindrone (or its acetate), levonorgestrel, MPA, dydrogesterone, and progesterone, respectively. The specific dose recommended also depends on whether the proges-

<table>
<thead>
<tr>
<th>Progestogen</th>
<th>Experimental</th>
<th>Based on dose</th>
</tr>
</thead>
<tbody>
<tr>
<td>Levonorgestrel</td>
<td>8</td>
<td>2–6.7 (0.15–0.5 mg)</td>
</tr>
<tr>
<td>Norethindrone</td>
<td>1</td>
<td>1 (1 mg)</td>
</tr>
<tr>
<td>MPA</td>
<td>0.1</td>
<td>0.1–0.4 (2.5–10 mg)</td>
</tr>
<tr>
<td>Dydrogesterone</td>
<td>0.02</td>
<td>0.05 (20 mg)</td>
</tr>
<tr>
<td>Progesterone</td>
<td>0.002</td>
<td>0.01–0.0033 (100–300 mg)</td>
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Potency values are relative to a value of 1 for norethindrone.
togen is given sequentially for 12–14 d/month or continuously as well as on the type of estrogen administered concurrently.

In contrast to animal experiments and clinical data, several researchers have done experiments in cell culture to investigate more directly the relative potency, efficacy, and biocharacter of progestogens via specific steroid receptors and on specific target genes. These strategies include the use of cell lines as models for cells in a particular target tissue relevant to HT side effects, cell lines deficient in other receptors with transient overexpression of the receptor under investigation, or the genetic engineering of cell lines to overexpress a particular receptor. However, very few studies have verified the specificity of the response by, for example, small interfering RNA knockdown of a particular receptor or using receptor-specific antagonists. Nevertheless, much valuable information has been obtained from these in vitro activity studies, including evidence for a lack of a class effect of progestogens.

C. Regulation of transcription by progestogens: genomic effects

1. Overview of mechanisms of ligand-dependent transcriptional regulation by steroid receptors

Ligand-activated steroid receptors directly regulate transcription of specific target genes by several genomic mechanisms that are conserved within the family of steroid receptors, although some mechanistic differences do occur. Regulation of transcription of mammalian genes generally involves dynamic, regulated steroid receptor-mediated recruitment of multiprotein complexes. These complexes include chromatin-remodeling proteins that shift nucleosomes, coactivators that acetylate histone proteins to open up chromatin, or corepressors that deacetylate histone proteins resulting in more compact chromatin. Also involved are several other proteins such as mediator complexes, the basal transcription machinery including RNA polymerase and associated factors, and enzymes that modify components of the complexes, including methylases and kinases (122, 123). Steroid receptors are key proteins in this process (99). In the absence of ligand binding, the PR and ER are located predominantly in the nucleus, whereas the AR, GR, and MR are located predominantly in the cytoplasm (44). There is also evidence that receptor isoforms display differential subcellular localization in the absence of ligand. For example, in endometrial cancer cells, the unliganded PR-A is predominantly located in the nucleus, whereas the unliganded PR-B is predominantly cytoplasmic (124), but both PRs are distributed in the nucleus and in the cytoplasm of several cell lines when overexpressed (125). The receptors are held in an inactive conformation by the presence of a protein complex of the heat-shock proteins (hsp) hsp90 and hsp70, immunophilins, and other proteins (126). The lipid-soluble steroid ligands diffuse passively across the plasma membrane and bind to the LBD of steroid receptors, inducing hyperphosphorylation, a conformational change in the receptor, changes in the composition of the protein complex, and nuclear translocation of the cytoplasmic receptors (44, 60, 127).

The genomic mechanisms whereby ligand-bound steroid receptors directly increase transcription of many target genes via direct DNA binding, or transactivation, involve binding of a receptor dimer to specific palindromic DNA sequences in promoters of target genes known as steroid-responsive elements (SREs). This results in formation of a multiprotein complex on the promoter via protein-protein interactions, including chromatin-remodeling proteins, coactivators, and components of the transcriptional machinery, in a dynamic, complex interplay of factors leading to an increase in transcription initiation (41, 44, 47, 128) (Fig. 7). Although each steroid receptor exhibits selectivity and a higher affinity for specific SRE sequences, the high degree of structural and functional conservation within the DBDs of steroid receptors allows most steroid receptors to bind, at least in vitro, to the same DNA response element (reviewed in Ref. 129). Thus, the progesterone response element (PRE) also binds the AR, GR, and MR (reviewed in Ref. 130).

Ligand-bound steroid receptors can also transrepress or directly and negatively regulate transcription via several genomic mechanisms, including direct DNA binding to negative SREs (131), or by protein-protein interaction and interference with other DNA-bound transcription factors such as nuclear factor-κB (NFκB) or activator protein-1 and CCAAT-enhancer-binding protein (41, 132–137) (Fig. 7). The latter mechanism is often referred to as a tethering mechanism, which can also result in an increase in transcription, depending on the transcription factors involved and promoter architecture (138). The details of these mechanisms are not well established for most members of the steroid receptor family but have been the focus of studies on GR actions due to their involvement in the antiinflammatory response (133, 135). All the members of the steroid receptor family have been shown to repress genes by antagonizing NFκB action (133, 135, 137, 139–142). In addition, the PR has been shown to increase transcription via tethering mechanisms involving interaction with specificity protein 1 and CCAAT/enhancer-binding protein (143–145).

Ligand-bound steroid receptors can thus lead to both increases or decreases in transcription and hence gene expression, via several direct genomic mechanisms where the outcome is cell and promoter dependent, depending on which cofactors are recruited by the receptor and the iden-
tivity of the specific ligand. Several lines of evidence show that, in general, an agonist bound to a receptor induces a conformational change that facilitates binding of coactivators, resulting in transcriptional activation due to their intrinsic histone acetylase activity, which makes the chromatin more accessible for recruitment of the basal transcription machinery and other transcription factors (146). Antagonists, on the other hand, are generally accepted to promote either the recruitment of corepressors, resulting in a decrease of transcription initiation via their histone deacetylase activity, reducing accessibility of DNA-binding sites for transcription factors, or a failure to recruit coactivators (147, 148). However, this general description is likely to be an oversimplification, because the spatial architecture and three-dimensional packaging of chromatin inside the nucleus, as revealed by new chromatin immunoprecipitation-sequencing technology, may well play a major role in nuclear receptor action (149, 150). Furthermore, tissue-specific steroid responses are determined by tissue-specific expression profiles of cofactors that affect the differential recruitment of coactivators vs. corepressors (146). Thus, it is almost impossible to predict the transcriptional response for a particular steroid ligand on a particular gene in a specific cell type,

Figure 7. Schematic diagram to illustrate differential genomic (nuclear) and nongenomic (extranuclear, cytoplasmic) actions of progestogens and endogenous steroid hormones. The two progestins, MPA and norethindrone, were chosen to illustrate the concept of differential actions compared with each other and progesterone. In genomic actions, all progestogens bind to the PR and act as agonists. MPA is a partial to full agonist for the GR and AR but has no significant activity via the MR or ER. However, norethindrone is a partial to full agonist for the AR but has no significant activity via the GR, MR, or ER. Progesterone is a weak agonist for the GR and AR, has no significant activity via the ER, and is a full antagonist for the MR. The two best-characterized genomic mechanisms for steroid receptors are illustrated. The first is transactivation by steroid receptor dimers binding directly to SREs in the promoters of target genes, followed by recruitment of coactivators and increased transcription. The second is transrepression via tethering of a steroid receptor monomer to other positively acting transcription factors, followed by recruitment of a corepressor and inhibition of transcription. Other complexes and higher-order effects on chromatin structure as discussed in the text are not depicted for simplicity. The depicted nongenomic or cytoplasmic actions include activation of various cytoplasmic targets by the classical nuclear steroid receptors or by membrane steroid receptors. Progesterone is a full agonist for the mPR, whereas MPA and norethindrone have no significant activity via mPR. Note that cytoplasmic actions can also lead to genomic actions by targeting of nuclear proteins such as transcription factors, cofactors, and chromatin proteins or even steroid receptors. Also depicted is the cross talk between the classical PR and other plasma membrane receptors (R) such as the epidermal growth factor receptor, as discussed in the text. Note that actions of progesterone as a weak GR or AR agonist are not depicted. ALD, aldosterone; CORT, cortisol; E2, estradiol; mER, membrane ER; NET, norethindrone; PROG, progesterone; SR, steroid receptor; TEST, testosterone; TF, transcription factor.
and these responses need to be determined experimentally.

The epigenome is emerging as a major regulator of cell-type-specific responses, regulating cell-type-specific gene expression profiles induced by nuclear receptors in response to ligands. The epigenome is dynamic and is a function of many factors including DNA methylation, higher-order chromatin structure such as chromatin looping, posttranslational modification of histone tails, and localization of histone variants (123). Nuclear receptor binding sites are present in enhancer elements that are brought into proximity with promoters by chromatin looping mechanisms that are programmed during cell lineage commitment (123) and are important regulatory elements in cell-specific gene expression (151, 152). In addition to cell-specific responses being mediated by epigenetic preprogramming of enhancers, nuclear receptors can also reprogram the epigenome in response to ligands (123). Nuclear receptors associate with many of the enzymes that modify histones and chromatin structure, such as the histone lysine demethylase, LSD1, which has been shown to associate with the AR (153) and to be important for nuclear receptor-mediated gene expression (154).

2. Differential effects of progestogens on specific gene expression via steroid receptors

Despite the general trends discussed above, progestogens exhibit cell-type-specific and gene-specific effects in particular models relevant to disease and side effects, due to multiple factors as discussed in previous sections. Thus, it is useful to consider what is known about the effects of different progestogens via different steroid receptors on transcription of specific target genes. Unfortunately, very few such detailed mechanistic studies have been performed or designed to compare effects of different progestogens or establish the receptors involved. However, those that have been performed shed useful insights into differential intracellular progestogen actions.

a. Effects via PR. Side effects associated with progestins in HT use include increased risk of breast cancer (155–157), cardiovascular complications such as strokes (1, 158, 159), effects on immune function (160–163), and neurological effects (164, 165). There is evidence that the dose and choice of progestin could determine risk outcome (166–169). However, surprisingly little is known about the molecular mechanisms, differential effects, and target genes of progestins acting via the PR in target tissues relevant to these side effects.

Much research has focused on the mechanism of action of progestogens in human breast cancer cell lines, where both pro- and antimitogenic effects have been ascribed to PR agonists (137). Some reports suggest that similar genomic effects occur with most progestins and progesterone via the PR on several target genes (104). For example, microarray analysis revealed that MPA and progesterone exhibit very similar qualitative expression profiles, with MPA being somewhat more efficacious, on endogenous PR-regulated genes in the human T47Dco breast cancer cell line expressing both the PR-A and PR-B isoforms (105). Interestingly, the same authors detected some cell-specific differences in breast cancer cell lines between the maximal responses and potencies of MPA compared with R5020 on a synthetic PRE-luciferase construct, most likely due to different relative concentrations of proteins other than the PR, although this was not established (105). However, consistent with its lower PR agonist potency (92), drospirenone has been shown to display weak effects compared with progesterone and other progestogens such as MPA, norethindrone acetate, levonorgestrel, and trimegestone on the transcriptional profile of PR-regulated gene expression in the PR-positive T47Dco breast cancer cell line (104). Some progestins such as norethindrone are implicated in increased proliferation and metastasis via an induction of vascular endothelial growth factor (VEGF) release into the media of cultured T47D breast cancer cells (106) by a mechanism involving transactivation via three functional PRE elements in the VEGF promoter (107). Some evidence also exists that progestogens play a role in the development of PR+ breast cancer by affecting the ability of cancer cells to invade the surrounding environment and interact with the extracellular environment. Progesterone, MPA, and drospirenone have also been implicated in PR-mediated increased breast cancer cell migration, with drospirenone being less potent than MPA (108), similar to the differential effects observed in the T47Dco cell line.

In contrast to results in breast cancer cell lines where most progestins appear to have similar qualitative effects on transcription of target genes compared with progesterone, results in endometrial cells suggest that some progestins may have opposite effects compared with progesterone. For example, MPA has been shown to repress expression of the chemokine regulated on activation, normal T cell expressed and secreted (RANTES) gene via the PR in cultured human endometrial stromal cells (170), whereas progesterone increased the expression of RANTES in primary endometrial T cells (171). As found in breast cancer cells, MPA appears to have similar genomic effects via the PR-A and PR-B in endometrial cells, as suggested by the finding that MPA increases VEGF synthetic promoter activity in Ishikawa endometrial adenocarcinoma cells (107) via both receptor isoforms.

Results in cell line models relevant to cardiovascular side effects also suggest different genomic effects of some progestins compared with progesterone. For example, unlike progesterone and Dienogest, it was found that MPA, norethindrone acetate, and levonorgestrel increase ex-
pression of two markers of vascular inflammation, intracellular adhesion molecule-1 (ICAM-1) and vascular cell adhesion molecule-1 (VCAM-1), in part via the PR (172). Consistent with this, studies on endothelial nitric oxide (NO) production, a marker for vasodilation (173), suggest differential actions of some progestins compared with progesterone. MPA was shown to have no effect on NO production in isolated human endothelial cells as well as in aortas from ovariectomized rats, unlike progesterone and drospirenone, which increased NO production, most likely via the PR (174, 175).

Similarly, studies in rat models suggest differential actions on brain mitochondrial function of MPA compared with other progestins and progesterone, which is of relevance to neurological health in premenopausal and postmenopausal women (169, 176). Unlike progesterone, MPA antagonizes estrogen up-regulation of brain mitochondrial function. Although the detailed mechanisms are unknown, these most likely involve differential steroid receptor-mediated changes in expression of key genes such as ATP synthase (169, 176).

The above evidence suggests that progestogens exhibit differential genomic effects in several cell models relevant to breast cancer and endometrial, cardiovascular, and brain function. However, the molecular mechanisms and occurrence of ligand-, cell-, isoform-, and promoter-specific effects of a range of progestogens remain to be further investigated in parallel in more physiologically relevant primary cell models. In particular, the contribution of off-target actions via steroid receptors other than the PR requires further investigation as a possible explanation for differential progestogen actions. In addition, some of the observed effects of progestogens on gene expression may occur by indirect genomic actions via the PR or other steroid receptors (42). A physiologically important example of indirect genomic effects on estrogenic activity of progestogens via the PR is the up-regulation by the PR due to its transactivation of the 17β-hydroxysteroid dehydrogenase type 2 gene, the product of which inactivates estradiol by converting it to estrone (7). In addition, progestogens exert indirect antiestrogenic effects in the endometrium by transrepression of the ER gene (7). These antiestrogenic actions of progestogens in the endometrium do not occur via binding of progestogens to the ER.

b. Effects via AR. Several earlier-generation progestins possess androgenic activity but not antiandrogenic activity, whereas most of the newer progestins possess antiandrogenic activity but no androgenic activity in animal models. However, the relative advantages of androgenic or antiandrogenic actions of progestogens in HT, as well as the extent to which these are mediated via direct genomic AR actions, are unclear. Nevertheless, there is evidence that off-target effects of progestins via the AR are likely to be relevant to cardiovascular function and breast cancer in HT users. The rationale for using progestins with antiandrogenic activity in HT is to improve the poor lipid profile of postmenopausal women, attributable to decreased levels of estrogen and SHBG; the resulting increased levels of free biologically active androgen are associated with decreased levels of high-density lipoprotein (HDL) cholesterol and increased levels of low-density lipoprotein cholesterol (177). The antiandrogenic metabolic effects of several progestins are, however, not ascribed to binding to the AR, but rather to a competitive inhibition of 5α-reductase activity, thereby decreasing the conversion of testosterone to the more active DHT (177).

Consistent with an apparent requirement for progestins to lack GR activity, most progestins exhibit no transcriptional activity via the GR, with the ex-
ception of MPA, which has a high affinity for GR and exhibits potent GR agonist or partial agonist activity, and gestodene, which exhibits less potent GR partial agonist activity (22, 41). Strong evidence for GR-mediated agonist activity of MPA was obtained by chromatin immunoprecipitation analysis that showed the recruitment of the GR to a glucocorticoid response element (GRE)-containing endogenous promoter in response to MPA but not to nor-ethindrone acetate or progesterone (183). Several lines of evidence suggest that MPA, which is widely used as an injectable contraceptive and in HT, and by implication to a lesser extent gestodene, have side effects on immune, cardiovascular, bone density, breast cancer, and neurological processes via direct GR-mediated effects on gene expression. Consistent with the immunosuppressive properties of glucocorticoid ligands acting genomically as agonists via the GR, MPA has been reported to inhibit proliferative responses to the T-cell mitogens, concanavalin A, and phytohemagglutinin (100); it also, together with estrogen, down-regulates release of the proinflammatory cytokines IL-2 and interferon-γ by phytohemagglutinin-stimulated peripheral blood mononuclear cells isolated from postmenopausal women using HT (163). This is further supported by the finding that MPA, but not progesterone, transpresses IL-2 transcription in isolated human monocytes (77). Also, consistent with a role for GR-mediated transrepression by MPA in osteoporosis, MPA has been shown to display glucocorticoid-like negative effects on bone density, unlike other progestins such as norethindrone and levonorgestrel (184, 185).

Extensive work in cell lines has identified several target genes and provided evidence for direct GR-mediated effects of MPA in their transactivation and transrepression relevant to MPA’s off-target side effects mentioned above. MPA has been shown to exhibit potent GR-mediated transcriptional activity on synthetic and endogenous GRE-, activator protein-1-, or NFκB-containing promoters, respectively, in several cell lines (73, 98, 99, 186–190). For example, of relevance to immune function, MPA has been shown to repress IL-6 and/or IL-8 mRNA and/or protein levels in the L929sA (98) and human thyroid cancer (KTC-2) cell lines (186) as well as RANTES mRNA levels in an ectocervical cell line (181). Given the central role of glucocorticoids and the GR in the immune and inflammatory response (191, 192), MPA and possibly gestodene are likely to exert side effects on other target genes involved in immune function. In contrast, dienogest, drospirenone, and trimegestone do not possess glucocorticoid activity via overexpressed GR and synthetic GRE-containing promoters (90, 95, 97). Other evidence suggests GR-mediated genomic effects of MPA via GRE-containing genes on kidney function, because MPA, but not progesterone, was shown to increase endogenous α-ENaC (α-subunit of epithelial Na channel) in serum and sgk1 (glucocorticoid-regulated kinase 1) mRNA levels in mouse cortical collecting duct cell lines (188). Similarly, GR-mediated transactivation is implicated in cardiovascular side effects for MPA and gestodene, which, unlike norethindrone and levonorgestrel, up-regulate proteolytically activatable thrombin receptor (PAR-1) mRNA in rat vascular smooth muscle primary cells to potentiate the vascular procoagulant effects of thrombin (193). Relevant to possible beneficial effects on cardiovascular function, MPA acting most likely via the GR down-regulates endothelial nitric oxide synthase (eNOS) mRNA expression and resulting nitric oxide levels in human umbilical vein endothelial cells, unlike levonorgestrel (190).

Strong evidence exists for an important role of the GR in determining the differential actions of progestogens in breast cancer (194) via genomic mechanisms (195). Progesterone and progestins demonstrate distinct but overlapping mRNA expression profiles in breast cancer cells (195, 196). Recent results suggest that MPA, acting via transactivation of the GR on target genes such as the fatty acid synthase gene, may promote tumorigenesis in normal cells and cancer progression in cancer cells, unlike progestosterone (195). However, MPA acting via the GR has also been implicated in playing a positive role in breast cancer via increasing expression of nucleoside diphosphate kinase A (metastasis suppressor) protein expression in metastatic human breast carcinoma cells by transactivation via GREs (130). Unlike progestins such as norethindrone, MPA acting via the GR is likely to have an immunosuppressive effect via repression of cytokine gene expression in both systemic and local endocervical immune function (Hapgood, J. P., Y. Govender, R. M. Ray, C. Avenant, and M. Tomasicchio, unpublished results). Because progestins such as norethindrone, levonorgestrel, dienogest, drospirenone, and trimegestone do not act via the GR, they are likely to exhibit very different side-effect profiles on the above-mentioned targets compared with MPA and, to a lesser extent, gestodene.

d. Effects via MR. Unlike most progestins, some such as drospirenone and trimegestone exhibit potent antimineralocorticoid properties in vitro via binding with relatively high affinity to the MR and acting as aldosterone antagonists (31, 90). Their development appears to be based on a strategy to mimic the antimineralocorticoid properties of progesterone and/or to prevent cardiovascular complications in postmenopausal women using estrogen/progestin treatment for HT (94–96). Estrogen can promote an increase in weight and blood pressure via its actions on the
renin-angiotensin-aldosterone system, leading to sodium and water retention (197). Interestingly, no changes were observed in blood pressure with progesterone administration to normotensive postmenopausal women, although a slight reduction in blood pressure was observed in hypertensive women (198). However, the anti-MR effects of progesterone may be relevant only when endogenous progestosterone concentrations are high, such as during the luteal phase of the menstrual cycle and pregnancy, because progesterone has a short half-life and is rapidly converted to metabolites without anti-MR activity (199). Given the established role of aldosterone in regulating blood pressure and cardiovascular function, as well as effects on renal inflammation and the central nervous system (70–72), it is likely that progestogens with anti-MR activity will exert biological effects on these processes, depending on their affinity for the MR and the concentration of progestogens and competing ligands. This antimineralocorticoid effect has been evident by a slight decrease in body weight and blood pressure in women using drospirenone, but not levonorgestrel, in combination with estrogen (200). This is consistent with a study in ovariectomized female rats treated with aldosterone and salt to induce renal injury, which showed that estradiol in combination with drospirenone did not increase sodium retention and blood pressure, unlike MPA (201). Very little is known about the target genes or precise MR-mediated genomic mechanisms of progestogens.

Consistent with their low affinity for the MR, progestogens like MPA and norethindrone do not display transactivation agonist activity via the expressed MR on a reporter gene in the COS-1 cell line, although both were able to weakly antagonize aldosterone-mediated transcription via the MR, albeit to a much lesser extent than progesterone (97). In vitro studies have also confirmed that drospirenone exhibits antagonist activity toward aldosterone and weak agonist activity in transactivation studies via the MR (90, 202). Drospirenone, like progesterone, has been shown to inhibit aldosterone-induced up-regulation of the adhesion molecule E-selectin, plasminogen activator inhibitor-1, and the chemokine monocyte chemoattractant protein-1 in human female aortic endothelial cells, consistent with MR-mediated antagonist activity toward transactivation (202). The effects of antagonism of MR-mediated transrepression by progestogens like drospirenone remain to be investigated. Because most progestogens do not act via the MR, they are likely to exhibit very different physiological effects compared with progestogens such as drospirenone with potent anti-MR activity in tissues containing MR.

e. Effects via ER. Although most studies report no direct binding to, or genomic actions by, progestogens via the ER, some reports (203, 204) but not others (78, 81, 82) suggest that both MPA and norethindrone acetate or norethindrone do bind to the ER. The metabolites of norethindrone, gestodene, and levonorgestrel, have, however, been reported to activate the ER (180, 205, 206). Metabolites of norethindrone appear to discriminate between ER isoforms, its 5α-reduced metabolite (3β,5α-tetrahydro-norethindrone) having been shown to selectively transactivate ERα at low concentrations but being ERβ agonistic at high concentrations (206). Although the physiological significance of binding and genomic actions of some progestogens or their metabolites via the ER are unclear, the ER does play an important indirect genomic role in the actions of progestogens because the estrogen-activated ER regulates expression of the PR gene and hence the response to progestogens (7).

In summary, besides affinity, other major determinants of differential progestogen actions via a particular steroid receptor are their potency and efficacy for a particular biological response. However, the relationship between the affinity, potency, efficacy, and biocharacter of a ligand is not straightforward or predictable (41, 99) and appears to depend on which ligand, promoter, or cell is involved. For example, two ligands may bind to a particular receptor with a similar affinity, but one may be an agonist and another an antagonist, depending on the particular receptor conformation induced by that ligand (146). For a particular ligand and receptor, the potency and efficacy are also gene specific; i.e., a progestogen may be a partial agonist on one promoter but a full agonist on another in the same cell (99, 181). These promoter-specific differences are also dependent on chromatin structure and the particular promoter architecture and, hence, assembled multiprotein complexes that differ for each promoter. Relatively little work has been done to investigate the relative effects on gene expression via different progestogens in different cells and to investigate the mechanisms and receptors involved. Results to date do show, however, that many target genes relevant to disease and side effects are indeed differentially regulated by progestogens, most likely due to differential extents of involvement of different steroid receptors and their isoforms. However, much more research needs to be done, in particular to compare different progestogens in parallel as well as determine involvement of receptors and their isoforms and cofactors, e.g., by small interfering RNA knockdown experiments or by using receptor-specific antagonists. Cell-type-specific responses to progestogens are also most likely regulated by cell-type-specific epigenomic factors, as has been shown for other nuclear receptor ligands (123). It remains to be investigated whether differential cell-specific effects of progestins acting via the same nuclear receptor may be
mediated by differential interaction of the liganded receptor with proteins involved in epigenetic preprogramming or reprogramming, such as histone methylases, demethylases, and chromatin-remodeling proteins.

D. Nongenomic effects of progestogens

Besides regulation of transcription via binding to intracellular steroid receptors by so-called nuclear or genomic mechanisms, progestogens have also been reported to result in a range of cytotoxic effects such as activation of kinase pathways. These nongenomic effects have been reported to occur via the classical cytotoxic and nuclear receptors as well as by plasma membrane-bound classical steroid receptors and via other novel membrane-bound receptor proteins (67, 207–210). Nongenomic signaling via a membrane-bound PR has been implicated in playing a role in brain signaling, oocyte maturation, and breast cancer (210). Progestogens acting via the PR have been reported to activate several kinase pathways such as Src, MAPK, phosphatidylinositol 3-kinase/protein kinase B, and human epidermal growth factor receptor 2 tyrosine kinase signaling cascades (211–214). For example, cytoplasmic nongenomic signaling has been reported to occur in breast cancer cells via the cytosolic PR-B, which activates c-src and MAPK as well as Wnt-1 and the epidermal growth factor receptor (144). MPA acting via the PR has been shown to activate the signal transducer and activator of transcription 3 pathway via a mechanism involving rapid, nongenomic tyrosine phosphorylation in breast cancer cells (214). Furthermore, the mechanism also involves nongenomic induction of human epidermal growth factor receptor 2 nuclear translocation (215) to stimulate late cell growth. In most cases, the effects of different progestogens on activation of kinases have not been investigated, with most researchers using either R5020 (144, 212, 215) or MPA (214) simply as a potent PR reference agonist. One study that compared the effects of MPA vs. progesterone on kinase activation found that whereas both MPA and progesterone activate the Erk MAPK, only progesterone resulted in Erk nuclear translocation (211), suggesting that more research needs to be done to investigate differential kinase activation by progestogens. Whether a membrane-associated PR is the same protein as the intracellular PR is controversial. Some studies have reported the possible involvement of a novel cell surface membrane PR (mPR), which is similar to G protein-coupled receptors and couples to a G protein (216). However, although this mPR was reported to bind progesterone, it did not bind to several progestins such as MPA and norethindrone (216–218).

Nongenomic actions of progestogens via the PR have been implicated in playing an important role in breast cancer. Work in human breast cancer cell lines shows that MPA induces cell proliferation by increasing cyclin D1 promoter activity via the PR-B isoform, but not PR-A (219). The mechanism appears to involve cytoplasmic activation by the PR-B of the phosphatidylinositol 3-kinase/protein kinase B/NFκB signaling cascade, resulting in activation of the cyclin D1 promoter, which does not contain PRE-related sequences (219).

The mechanisms and physiological significance of nongenomic signaling by different progestogens via the PR isoforms remain to be further investigated and could potentially be relevant to differential side effects. Off-target differential nongenomic signaling by progestogens via steroid receptors other than the PR may also be physiologically relevant. These may, for example, be involved in mediating differential actions of progestogens in brain mitochondrial function (169, 176). Interestingly nongenomic actions mediated via the GR and ER have been reported to be involved in neuroprotection (220, 221), whereas nongenomic actions via the AR are likely to be involved in spermatogenesis (222). For the MR, nongenomic actions have been implicated in several physiological processes including brain signaling, endothelial dysfunction, and inflammation (209). Given the differential binding affinities and genomic actions of most progestogens via the intracellular classical ER, AR, and GR, it is likely that progestogens also exhibit differential nongenomic actions via these receptors when membrane bound or possibly via other novel membrane-bound steroid receptors.

Clinical Effects of Progestogens in Postmenopausal Hormone Therapy

Estrogen therapy is effective in treating climacteric symptoms and in the prevention of menopausal osteoporosis. The addition of progestogen in combination or sequential regimens has risks and benefits with regard to the endometrium, breast, cardiovascular system, bone, and brain.

A. Effects on the endometrium

As stated earlier, a progestogen is defined as a substance that transforms an endometrium primed by estrogen into a secretory endometrium. Progestogens are therefore used in HT in women with uteri to prevent the endometrial hyperplasia and endometrial cancer that may result from the use of estrogen only, often referred to as unopposed estrogen. Progestogens exert their protective effects by decreasing nuclear mitotic activity induced by estrogens and by increasing 17β-hydroxysteroid dehydrogenase type 2 activity that converts estradiol to the biologically less potent estrone.
The development of endometrial hyperstimulation with estrogen use increases with higher doses and duration of unopposed estrogen (223–225). Most recently, the Million Women Study (226) estimated the number of endometrial cancers per 1000 women in 5 yr to be 3.0 (2.8–3.2) without HT, 4.9 (3.5–7.5) with unopposed estrogen therapy, and 2.0 (1.5–2.6) with combined estrogen and progestin therapy; expressed in terms of relative risk (RR), HT never-users had a RR of 1.0, unopposed estrogen users had a RR of 1.45 [95% confidence interval (CI) = 1.02–2.06; \( P = 0.04 \)] and combined continuous estrogen with progestogen users had the lowest RR of 0.71 (95% CI = 0.56–0.90; \( P = 0.005 \)). Examination of the participants’ endometrial histology in the Postmenopausal Estrogen/Progestin Interventions (PEPI) Trial showed consistent endometrial protection across all treatment groups, comprising conjugated equine estrogens (CEEs) combined with MPA given either continuously (0.625 mg CEE/2.5 mg MPA) or sequentially (0.625 mg CEE/10 mg MPA), or 0.625 mg CEE plus 200 mg micronized progesterone, compared with placebo (the placebo group had one case of endometrial adenocarcinoma) (227). However, the European Prospective Investigation into Cancer and Nutrition (\( n = 115,474 \)) noted an increased endometrial cancer risk in all hormone users including treatment with estrogen only [39 cases, hazard ratio (HR) = 2.52; 95% CI = 1.77–3.57], estrogen plus progestin (121 cases, HR = 1.41; 95% CI = 1.08–1.83), and estrogen plus micronized progestosterone (26 cases, HR = 2.42; 95% CI = 1.53–3.83) (228). The authors stated that risks differed according to regimen and type of progestin used and that because the micronized progestosterone findings are based on small numbers, they recommended further study. This study also found that continuous regimens may better reduce the risk of endometrial cancer; the group receiving continuous treatment had only three cases of endometrial cancer (HR = 0.24; 95% CI = 0.08–0.77), whereas the sequential group had 50 cases (HR = 1.52; 95% CI = 1.00–2.29) (228). In light of these data, continuous daily combined estrogen/progestogen treatment as well as daily estrogen with sequential progestin (for 12–14 d/month), has been advocated to decrease estrogen-stimulated risk of endometrial adenocarcinoma and its precursor hyperplasia. Typical daily dosages of progestogens used for endometrial protection in women using estrogen therapy are shown in Table 6.

Progestogens are available in various formulations that can be prescribed for endometrial protection. Progesterone vaginal gel (4%) used biweekly for 1 yr together with a transdermal estradiol patch releasing 50 \( \mu g/d \) estradiol resulted in atrophic endometrium being observed on biopsy in all subjects (229). Progesterone capsules (100 mg) administered orally every other day combined with a transdermal patch releasing 50 \( \mu g \) estradiol per day for a duration of 3 yr also resulted in endometrial biopsies showing atrophy in all women (230). Additionally, the levonorgestrel intrauterine system has been studied for endometrial protection when systemic estrogen is used. There was no endometrial hyperplasia in three studies using the levonorgestrel intrauterine system in postmenopausal women receiving 1.25 \( mg/d \) oral CEE (231), 1.5 \( mg/d \) estradiol gel (232), and either a 50-\( \mu g \) estradiol patch or 2 \( mg \) oral estradiol valerate (233), each for a duration of 5 yr.

### B. Effects on the breast

Although *in vitro* experiments cannot replace clinical trials, they are useful to explore possible differences between substances tested in the same model, which can then be confirmed in clinical studies. There are numerous experimental data available on the effect of progestogens on proliferation of normal and cancerous breast epithelial cells, but only a limited number of experiments have been carried out testing multiple progestogens in the same cell model. Of particular interest is an *in vitro* study in which the effects of progesterone, MPA, chlormadinone acetate, norethindrone, levonorgestrel, gestodene, and dienogest on proliferation and apoptosis of normal breast epithelial cells were tested at various concentrations, and the ratio of apoptosis to proliferation was compared (234). MCF-10A, a human, nontumorigenic, estrogen- and PR-negative breast epithelial cell line, was used with a mixture of growth factors to stimulate the cells. In combination with growth factors, the apoptosis/proliferation ratio was reduced significantly by MPA and chlormadinone acetate, favoring a proliferative effect. MPA produced as much as

### TABLE 6. Typical daily progestogen doses used for endometrial protection in hormone therapy

<table>
<thead>
<tr>
<th>Progestogen</th>
<th>Dose (mg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Progesterone</td>
<td>200–300</td>
</tr>
<tr>
<td>MPA</td>
<td>2.5–10.0</td>
</tr>
<tr>
<td>Chlormadinone acetate</td>
<td>10</td>
</tr>
<tr>
<td>Cyproterone acetate</td>
<td>1</td>
</tr>
<tr>
<td>Dydrogesterone</td>
<td>5–10</td>
</tr>
<tr>
<td>Nomegestrol acetate</td>
<td>5–10</td>
</tr>
<tr>
<td>Promegetone</td>
<td>0.25–0.5</td>
</tr>
<tr>
<td>Trimegestone</td>
<td>0.5</td>
</tr>
<tr>
<td>Norethindrone acetate</td>
<td>0.5–1.0</td>
</tr>
<tr>
<td>Levonorgestrel</td>
<td>0.075</td>
</tr>
<tr>
<td>Desogestrel</td>
<td>0.075</td>
</tr>
<tr>
<td>Norgestimate</td>
<td>0.09</td>
</tr>
<tr>
<td>Gestodene</td>
<td>0.05</td>
</tr>
<tr>
<td>Dienogest</td>
<td>3–4</td>
</tr>
<tr>
<td>Drosperone</td>
<td>2</td>
</tr>
</tbody>
</table>

Values are based on Ref. 6.
a 4-fold reduction in the ratio. The other progestogens had no significant effect. The results suggest that the progestogens differ in their ability to induce proliferation or inhibit the growth of normal human breast epithelial cells. However, these results should be interpreted with caution because MCF-10A cells lack PRs and ERs and have a myoepithelial phenotype. This differs from the normal breast, which contains ER/PR-positive luminal cells that are likely the primary point of response to HT.

Information about the risk of a clinical breast cancer diagnosis associated with isolated progestogen use in postmenopausal women is theoretical in that there are few clinical indications other than HT, where it is used solely for endometrial protection in nonhysterectomized women. In premenopausal women using progestogens in hormonal contraception, clinical breast data on the progestogen-only effect is scant because the reproductive age group of women users has a low baseline risk of breast cancer. The result of adding a progestogen to estrogen in HT creates a more complex and confusing situation because there is no consensus on the background effect of estrogen itself as well as the effect of its dose, delivery system, treatment time, and the patient’s background medical and lifestyle attributes. Publication of the WHI study in 2002 (1) and subsequent analyses of data have generated an intense interest in the issue of independent risk from the progestogen when added to estrogen therapy, specifically regarding breast cancer. As a result, epidemiological studies are moving away from investigating estrogen alone compared with estrogen combined with any progestogen and instead comparing estrogen alone with estrogen combined with specific unique progestogens.

The WHI CEE/MPA study arm was stopped prematurely at 5.2 yr because of increased risk of breast cancer with a HR of 1.26 (95% CI = 1.0–1.6) (1) after an average of 7.1 yr. Although there was no increased risk of breast cancer diagnosis of invasive breast cancer in the estrogen-only arm, CEE compared with placebo (HR = 0.80; 95% CI = 0.62–1.04; P = 0.09), there was little immediate follow-up on this different outcome in the two arms (235). The WHI CEE/MPA study arm remained the focus of analyses because the cancers were more advanced and more commonly node-positive (81 cases, 30.7%, vs. 43 cases, 16.2%, respectively; HR = 1.96; 95% CI = 1.23–2.58).

Breast cancer mortality appeared to be greater in the CEE/MPA group (25 deaths, 0.03% per year) compared with the placebo group (12 deaths, 0.01% per year) (236). It was noted that there was increased mammographic density that interfered with mammographic detection, resulting in diagnostic delay as well as diagnosis of breast cancers at a more advanced stage (237).

Mammographic density is a breast cancer risk factor and is associated with increased risks for hyperplasia, atypical hyperplasia, and ductal carcinoma in situ. The earlier PEPI trial found greater breast density over 12 months regardless of whether the regimen was continuous (0.625 mg CEE/2.5 mg MPA) or whether it was sequential (0.625 mg CEE/10 mg MPA or CEE/200 mg micronized progesterone) (238). When HT was discontinued, mammographic density quickly returned to normal levels. The impact of dose was then studied in the combination regimens. In a study evaluating ultra-low-dose HT (0.5 mg estradiol/0.25 mg norethindrone acetate or 0.5 mg estradiol/0.1 mg norethindrone acetate), no change was found in breast density after 6 months of use (239).

Besides the WHI results, additional epidemiological data on postmenopausal HT have consistently reported that the addition of any progestin to estrogen increases the risk of breast cancer diagnosis compared with estrogen alone (240–245). A case-control study (n = 33,000) evaluating breast cancer risk from 1996–2006 in an insurance database population of women aged 50–64 yr (246) reported an odds ratio of 0.96 (95% CI = 0.88–1.06) for developing breast cancer in women using estrogen-only therapy, whereas in those using estrogen/progestin HT, the odds ratio was 1.44 (95% CI = 1.31–1.58). Another study evaluated pooled data from six mammographic registries (n = 373,265); HT users with and without hysterectomy were recruited and followed prospectively, noting the intervals between mammograms (247). Use of estrogen and progestin for greater than 5 yr was associated with a greater risk of breast cancer diagnosis (RR = 1.49; 95% CI = 1.36–1.63) compared with nonusers. The increased risk was not observed either in estrogen plus progestin users who had used HT for less than 5 yr (RR = 0.83; 95% CI = 0.73–0.98) or in estrogen-only users irrespective of duration of use whether less than 5 yr (RR = 0.86; 95% CI = 0.71–1.03) or more than 5 yr (RR = 0.92; 95% CI = 0.84–1.00) (247).

A 2006 meta-analysis of studies to assess evidence for a link between postmenopausal HT and risk of breast cancer diagnosis (248) found an average RR of 0.79 (95% CI = 0.61–1.02) for invasive breast cancer diagnosis with estrogen use and of 1.24 (95% CI = 1.02–1.50) with estrogen-progestin use in four randomized trials, whereas epidemiological studies reported a RR of 1.18 (95% CI = 1.01–1.38) with estrogen alone and 1.70 (95% CI = 1.36–2.17) with estrogen plus progestin (248). Hormone use and breast cancer histology was also evaluated in a prospective cohort (n = 67,754) (249); in the estrogen and progestin group, both ductal cancer (RR = 1.75; 95% CI = 1.59–2.01) and lobular cancer (RR = 2.12; 95% CI = 1.62–2.77) diagnoses were increased compared with the estrogen-alone group, in which
there was no increase in the diagnosis of ductal cancer (RR = 0.99; 95% CI = 0.84–1.17) or lobular cancer (RR = 1.13; 95% CI = 0.94–1.78) (249).

A recent WHI analysis (250) has reviewed the risk of breast cancer diagnosis in the CEE-alone study arm. After a median follow-up of 11.8 (9.1–12.9) yr with estrogen use for 5.9 (2.5–7.3) yr, there was a lower incidence of breast cancer (151 cases, 0.27% per year) compared with placebo (199 cases, 0.35% per year; HR = 0.77; 95% CI = 0.62–0.95; P = 0.02). In the estrogen-only arm, mortality per year from breast cancer (96 deaths, 0.009%) was less than that seen in controls who did not use HT (16 deaths, 0.024%), and in fact, fewer women in the estrogen-alone group than the control group died from any cause.

A current review (251) summarizes the conceptual change since 2002, based on WHI CEE and CEE/MPA data, in light of the fact that epidemiological studies consistently show estrogen when combined with progestin carries a different breast cancer risk compared with estrogen alone when used for 5 yr. The review reiterates the conclusions noted in previous citations with regard to CEE/MPA risks and states that estrogen-alone use reduces breast cancer risk and does not substantially interfere with breast cancer detection by mammography.

Clarification of the differential effects of different progestogens combined with estrogen is limited because few epidemiological studies have had sufficient sample sizes and/or accurate information to assess breast cancer risk in relation to different types and routes of estrogen administration associated with different types of progestogens. However, the observational Million Women Study (155, 252) found no significant differences in risk of breast cancer diagnosis between estrogen alone (RR = 1.30; 95% CI = 1.22–1.38) and estrogen combined with various progestins including tibolone (RR = 1.45; 95% CI = 1.25–1.67), estrogen/norethindrone (RR = 1.53; 95% CI = 1.35–1.75), estrogen/MPA (RR = 1.60; 95% CI = 1.33–1.93), and estrogen/norgestrel/levonorgestrel (RR = 1.97; 95% CI = 1.74–2.33). Moreover, until recently, micronized progesterone has not been included in such studies.

That changed with the publication of results from the French E3N Cohort Study (253), which assessed and compared the association between different HT regimens and breast cancer risk in 80,377 postmenopausal women, whose mean age was 53.1 yr. The women were followed up for an average of 8.1 postmenopausal years, during which they completed self-administered biannual questionnaires addressing medical history, menopausal status, and lifestyle characteristics. A total of 2354 cases of invasive breast cancer cases were identified primarily from the self-reports, 95.3% of which were confirmed by pathology reports. Information on lifetime use of hormonal treatments was also obtained from the questionnaires. The women were given a booklet listing the hormonal treatments marketed in France, complete with color photographs and products, to help them remember what preparations they had taken. Seventy percent of the women had used HT for a mean duration of 7 yr.

When RRs of invasive breast cancer associated with the most frequently used HTs were compared with never-used HT, RRs varied significantly between the different progestogens for any given route of estrogen administration (oral or transdermal). Estrogen-progesterone and estrogen-dydrogesterone combinations were associated with no or slight and nonsignificant increases in risk, whereas all other estrogen/progestogen combinations showed substantially increased risks, most of which were statistically significant but did not differ significantly between preparations. Other than progesterone and dydrogesterone, the progestogens included medrogestone, chlormadinone acetate, cyproterone acetate, promegestone, nomegestrol acetate, norethindrone acetate, and MPA. The latter two progestins were combined with oral estrogen compared with all the other progestogens, which were combined with transdermal estrogen. Thus, the estrogen delivery system was also a differential factor in the study (253).

Because of those findings, subsequent statistical evaluations included separate estimates for HTs containing progesterone or dydrogesterone, but the other progestogens were grouped together. When RRs for invasive breast cancer associated with type of HT and duration of exposure were compared with never-used HT, women in the estrogen-alone and estrogen/other progestogens groups had a significantly increased breast cancer risk, with RRs of 1.29 (95% CI = 1.02–1.65) and 1.69 (95% CI = 1.50–1.91), respectively. In contrast, estrogen-progesterone was associated with a RR of 1.00 (95% CI = 0.83–1.22) and estrogen-dydrogesterone with a RR of 1.16 (95% CI = 0.94–1.43). Estrogen-alone, estrogen-progesterone, and estrogen-dydrogesterone were associated with breast cancer risks that did not differ significantly from one another but were all significantly lower than the RR of estrogen-other progestogens (253).

Estrogen itself remains controversial regarding breast cancer risk. However, the epidemiological studies clearly delineate a different risk of diagnosis of breast cancer in estrogen-alone vs. estrogen plus progestogen treatments. Although the return to baseline risk in former HT users supports a promotional effect rather than an initiating effect of estrogen with progestogen in the risk of breast cancer diagnosis (254), the data do not support such a promotional effect from estrogen alone. It is only when the progestogen is added to the estrogen-primed breast tissue
that there is an increase in diagnosis of breast cancer. The emerging clinical epidemiological data support the hypothesis that progestogens are not a uniform class and that progesterone and progestins have different effects, with distinctive impacts on the risk of breast cancer diagnosis in menopausal women using HT.

### C. Effects on the cardiovascular system

The widespread prescription of HT to postmenopausal women was historically, in addition to relieving menopausal symptoms, intended to protect women from cardiovascular disease. However, results of the larger trials to test the benefits of HT in reducing cardiovascular events were disappointing. The 6.8-yr follow-up report on the Heart and Estrogen/Progestin Replacement Study found no significant reduction in primary or secondary coronary heart disease (CHD) events in the CEE/MPA group vs. the placebo group (253). Manson and co-workers (256) reported an increased risk of cardiovascular disease with CEE/MPA in the WHI trial, noting that this was most apparent only during the first year of use. Additional analysis of the WHI study concluded that women who initiated HT closer to menopause tended to have reduced CHD event risk vs. women more distant from menopause (257). Also, women who initiated therapy at a younger age had a lower CHD event risk compared with women who initiated therapy at an older age. Short-term use of HT in the immediate postmenopausal years has been advocated to protect against cardiovascular events in the long term, whereas initiating HT is not recommended for older women who are already at higher risk of cardiovascular disease (258).

Further examinations of those WHI study trends demonstrate a differential CHD event risk between the CEE/MPA group and the CEE-alone group (257). For example, women not stratified by age, who initiated HT within 10 yr of menopause in the CEE-alone group had decreased CHD events (HR = 0.48; 95% CI = 0.2–1.17) compared with the CEE/MPA group (HR = 0.88; 95% CI = 0.54–1.43). Also, the decrease in CHD events in the 50- to 59-yr group was more pronounced in the CEE-alone group (HR = 0.63; 95% CI = 0.36–1.09) when compared with the CEE/MPA group (HR = 1.29; 95% CI = 0.79–2.12). The decreased CHD events in the CEE arm vs. the CEE/MPA arm in these two patient groups suggest that it is the CEE/MPA that has the adverse impact. Therefore, there is a potential for cardiovascular protection with estrogen alone and perhaps by estrogen combined with other progestogens. This potential should be further explored.

Much has been written since the Heart and Estrogen/Progestin Replacement Study and WHI results were published about the cardiovascular implications of HT, largely concentrating on the type and route of administration of the estrogenic component because of its importance in modulating cardiovascular risk. Estrogen treatment improves lipid profiles and insulin sensitivity and has beneficial effects on mitigating central weight gain in menopausal women. Unfortunately, there are few long-term clinical studies comparing different progestogens used in HT with respect to cardiovascular outcomes. However, some aspects of potential cardiovascular risk have been examined, namely effects on lipids, vascular function/blood pressure, inflammation, thrombosis, and carbohydrate metabolism.

The most common comparison has been between progesterone and MPA, and Hermsmeyer et al. (259) have cautioned against a negative view of HT for cardiovascular protection based on only the results of trials involving MPA. When oral micronized progesterone was used in one group in the PEPI study in place of MPA, this group had significantly higher HDL cholesterol levels than the MPA group, indicating a more favorable effect on blood lipids (260), although there is no evidence that this improved cardiovascular outcomes. A small study of 18 women showed that progesterone vaginal gel produced an increase in exercise tolerance in postmenopausal women with coronary artery disease or previous myocardial infarction who were being treated with estradiol, whereas MPA did not, compared with estradiol alone (261). Primate studies have demonstrated a marked adverse effect of MPA on coronary artery hyperreactivity that is the opposite of the protective effect seen with progesterone (262). Furthermore, MPA, but not progesterone, negated the coronary vasospasm protective effects of estradiol, shown by measuring intracellular calcium and protein kinase C signals (263, 264), and progesterone reduced coronary hyperreactivity even in the presence of atherosclerosis in oophorectomized rhesus monkeys (265). Adverse effects on carbohydrate metabolism, namely higher fasting glucose and insulin levels, and higher insulin responses to glucose challenge have also been demonstrated in oophorectomized cynomolgus monkeys receiving CEE plus MPA compared with CEE alone or no HT (266).

A review of the effects of progestins on cardiovascular risk markers by Sitruk-Ware (159) showed that progesterone and its 19-norprogesterone derivatives, which have no androgenic effects, did not adversely impact the beneficial effects of estrogens on the lipid profile, notably in their ability to increase HDL cholesterol levels, whereas those with androgenic properties, namely the 19-nortestosterone derivatives and some 17-hydroxyprogesterone derivatives, including MPA, have shown negative effects on lipids. This observation has been confirmed in more recent studies, e.g. a comparison of trimegestone (either
inflammation that is implicated in the development of ath-

erosclerosis, whereas CEE plus nomegestrol acetate re-

duced high-sensitivity C-reactive protein levels in a ran-

donized study in postmenopausal women (273); all HT
groups showed a decline in homocysteine levels. On the
other hand, a small, randomized comparison of MPA or
oral micronized progesterone combined with CEE in 20
postmenopausal women found similar improvements in
both groups with respect to flow-mediated dilator re-
sponse to hyperemia as well as similar effects on markers
of inflammation, hemostasis, and fibrinolysis (274). Also,
a comparison of cyproterone acetate or MPA in 26 post-
menopausal women receiving estradiol valerate (275)
found that both progestins attenuated the beneficial ef-

fects of the estrogen on nitric oxide release, the mechanism
by which estrogen is thought to exert its vascular endo-

thelial effects.

Another advantage of the less androgenic progestins is
their more positive impact on the hemostatic system. In a
multicenter study of 186 postmenopausal women com-
paring estradiol plus either dydrogesterone or trimeges-
tone for 6 months (270), a decrease in protein C activity
and an increase in plasmin-antiplasmin complex were seen
in the trimegestone group. This suggested an enhanced
fibrinolytic response that could translate to a reduced risk
of thrombosis and consequent reduced risk of stroke or
myocardial infarction. Recent clinical studies outline the
contribution of the progestogen component of HT to
thrombotic risk. The French Study of Norpregnanes on
Coagulation compared hemostatic parameters with no
HT, transdermal estradiol plus micronized progesterone,
and transdermal estradiol plus norpregnane derivatives
(nomegestrol acetate or promegestone) (271). Thrombin
generation in the presence or absence of activated protein
C showed activated protein C resistance and therefore in-
creased thrombotic potential in the norpregnane group
but not in the micronized progesterone group compared
with no HT. Also, data on thromboembolism incidence
after an average follow-up of 10.1 yr in the E3N Cohort
Study of 80,308 postmenopausal women (272) showed
significantly increased thrombotic risk with norpregnanes
(HR = 1.8) compared with progesterone (HR = 0.9),
pregnanes (HR = 1.3), and 19-nortestosterone derivatives
(HR = 1.4).

CEE plus MPA or CEE alone was also found to increase
levels of high-sensitivity C-reactive protein, a marker of
inflammation that is implicated in the development of ath-

omy. There was no significant association of VTE with micronized progesterone and norpregnane derivatives but a 4-fold increased risk of VTE when norpregnane derivatives were used in combination HT. Progesterone derivatives included dydrogester-

one, medrogestone, chlormadinone acetate, cyproterone
acetate, and MPA, whereas the norpregnane derivatives were either nomegestrol acetate or promegestone. Studies of combined OCs have indicated that third-generation progestins (desogestrel or gestodene) may carry a greater VTE risk than OCs containing levonorgestrel (281). Although a recent review indicated that the newer progestin drospirenone did not appear to show an increased VTE risk compared with other progestins when used in combined OCs (282), two new case-control studies have found an increased VTE risk with drospirenone compared with levonorgestrel (283, 284). Also, a Danish cohort study showed that OCs containing levonorgestrel were associated with a 3-fold increase in risk for VTE compared with nonusers of OCs, whereas users of OCs containing desogestrel, gestodene, or drospirenone had a 6- to 7-fold increase in VTE risk compared with nonusers (285), i.e. at least twice the risk for levonorgestrel.

Although progestins have differing effects on aspects of cardiovascular risk, in general, those more similar to progesterone have been associated with a lower impact than the more androgenic progestins on the beneficial effects of concomitant estrogen therapy. However, the limited number of long-term clinical studies makes it difficult to extrapolate the short-term effects on various markers of cardiovascular risk to long-term cardiovascular morbidity.

D. Effects on the brain

Progesterone has important functions in the nervous system and has been classified as a neurosteroid. Endogenously, progesterone is synthesized de novo from cholesterol in the brain, spinal cord, and peripheral nerves, and its actions may be mediated by local metabolism to allopregnanolone (286, 287). Recent clinical studies have demonstrated significant neuroprotective activity of high-dose progesterone treatment in subjects with traumatic brain injury (288, 289), leading to additional phase III trials that are ongoing. Progesterone is a promising therapy for acute neuroprotection in brain injuries and neurodegenerative diseases, probably via multiple physiological mechanisms (290, 291).

Whether or not the neuroprotective benefits of progesterone can also be achieved by progestins has not been determined clinically (292). However, rat studies have demonstrated that the neuroprotective effects of progesterone are not seen with MPA (293, 294). In a model of estrogen-induced neuroprotection involving assessment of the effects of glutamate toxicity in rat hippocampal neuron cultures, both progesterone and 19-norprogesterone showed neuroprotection, either alone or in combination with estradiol, whereas MPA was not neuroprotective and even blocked the neuroprotective activity of estradiol in this model (295). The same group assessed the impact of progesterone and MPA on the excitotoxic glutamate-induced rise in intracellular calcium levels, a neurotoxic effect, which was attenuated by both progesterone and estradiol. MPA did not have an effect when administered alone to the hippocampal neurons, but when administered together with estradiol, MPA completely antagonized the attenuation by estradiol of the rise in intracellular calcium (296). The MAPK cascade is a mechanism involved in this estrogen-mediated neuroprotection, and the authors showed that MPA, but not progesterone, blocked the estradiol-induced nuclear translocation of extracellular receptor kinase, which is required for calcium regulation. Furthermore, when this group looked at survival of hippocampal neurons exposed to crystalline MPA or a pharmaceutical formulation containing MPA (Depo-Provera) in that model, both showed a lack of neuroprotective efficacy; also, medroxyprogesterone (without the acetate group) was as ineffective as MPA (297).

A study of acute MPA administration to brain-injured rats showed a dose-related reduction in cerebral edema but no improvement in performance on a spatial learning task, indicating some beneficial antiinflammatory effects but no functional improvement compared with progesterone (298). Thus, the differences between effects of progesterone and progestins that have been observed in other tissues may also pertain to neurobiological effects (287).

In the WHI memory study, greater brain atrophy, assessed by a reduction in hippocampal volume on brain magnetic resonance imaging, was seen with CEE either with or without MPA compared with the placebo group, although this was most apparent in women with cognitive defects before initiating HT (299).

A review of the effects of progesterone on the nervous system by Schumacher and co-workers (300) notes that it is a mistake to consider progestogens as a single class, because progestins have some very different properties from those of natural progesterone. The authors also note that the adverse clinical effects of some progestins should not discourage the use of natural progesterone or the development of new, safer progestins for use in postmenopausal HT to reverse age-dependent dysfunction of the nervous system.

E. Effects on bone

Estrogens are well known to prevent bone loss because of their physiological inhibitory effect on bone resorption through the osteoclasts. According to meta-analyses, randomized controlled trials of menopausal HTs consistently indicate improved bone density with estrogen use (301, 302). Norethindrone acetate has been shown to prevent bone resorption in postmenopausal women without added estrogen (303). High doses of MPA cause partial
reduction in bone resorption (304). Cyclic MPA has been found to increase bone mineral density in premenopausal women with anovulatory or short luteal-phase menstrual cycles (305); however, clinical investigations of the effect of progesterone have been inconclusive (306). Other progestins have not been well studied. Thus, there is limited evidence for an independent effect of progestogens on bone.

VIII. Conclusions

This review compared progestogens with respect to their chemical structures, structure-function relationships, metabolism, pharmacokinetic parameters, intracellular mechanisms, potency, efficacy, and biological and clinical effects. The chemical structures of progestogens vary widely. Some are structurally related to progesterone, others to testosterone, and one to spironolactone. Progestogens also differ in their metabolism and pharmacokinetic profiles. Some are prodrugs and require transformation to active forms, and they have wide differences in their bioavailabilities and half-lives. Although progestogens are designed to be potent and high-affinity PR agonists that mimic the biological actions of the natural ligand, progesterone, many of them bind to other members of the steroid receptor family, which include the AR, GR, and MR. Furthermore, they exhibit considerable variation in their binding affinities, potencies, and efficacies as well as the resulting extent of agonist, partial agonist, or antagonist responses via these receptors. All these differences are most likely major determinants of differential actions and the lack of a class effect of progestogens. Relative concentrations of free progesterone and competing endogenous ligands reaching the target cell would depend on relative affinities for serum binding globulins in the blood (87, 307) as well as half-life, metabolism, route, and method of administration and dosage (5, 7, 308–310). Once inside the cell, the fractional occupancy of a particular receptor, and hence the relative response via that receptor, would depend on concentrations of metabolizing enzymes in the target cell as well as relative intracellular concentrations of steroids and their relative affinities for different binding proteins.

Another major determinant of differential actions by progestogens is most likely the cell-specific concentration of proteins that affect the final biological response. These include variations in concentration and isoform type of classical intracellular and membrane-bound steroid receptors, nonsteroidal receptors that cross talk with the steroid receptor pathway, steroid-metabolizing enzymes, and all interacting partners in the various steps leading to the progestogen-induced biological response. For example, the effects of PR ligands on proteasomal-mediated turnover of coactivators like steroid receptor coactivator-1 are likely to affect progestogen genomic responses (311). Cell-specific effects can be dramatic; some steroid receptor ligands can even switch from agonist to antagonist, depending on the milieu of cell-specific cofactors (312), whereas a particular progestogen can act either as an agonist, a partial agonist, or an antagonist, dependent solely on steroid receptor concentration (187). The relative tissue-specific distribution and variable levels of steroid receptors are likely to play a major role in differential progestogen actions and side effects, given the variation in affinities of progestogens for different receptors.

The explicit clinical effects of progestogens are difficult to determine. In postmenopausal women, there is scant information on the clinical effects of progestogens alone, because progestogens are generally prescribed when estrogen is being used to prevent endometrial cancer associated with unopposed estrogen. Until the WHI trial, progestogens did not have a major role in the general HT discussion because they were prescribed only for endometrial safety should estrogen be prescribed. There was minimal concern for their contribution to the risk/benefit profile. The WHI findings have highlighted the need to understand the clinical effects of progestogens as well as those of estrogen. The differential clinical outcomes in the epidemiological studies of estrogen alone compared with combined estrogen and progestogen regimens have generated questions regarding progestogens and the recognition that they have potentially distinctive risk/benefit profiles when combined with estrogen. However, the clinical outcome data available are confusing due to the use of different estrogens and progestogens, different doses, delivery systems, and treatment regimens, and different risk attributes for the users. Post-WHI epidemiology is focusing on comparing the clinical outcomes of estrogen alone with estrogen combined with different specific progestogens.

In breast and cardiovascular studies, the outcome data show different RRs and lack of a class effect of progestogens. Although the Million Women Study shows that “benefits for endometrial cancer associated with continuous combined progesterin therapy may be outweighed by risks for breast cancer, which is adversely affected by the therapy” (252), only progestins, and not progesterone, were evaluated in the study. In the recent French E3N Cohort Study, estrogen plus progesterone or dydrogesterone regimens were associated with decreased invasive breast cancer risk compared with regimens consisting of estrogen combined with other progestogens. Although the endometrial protection from progestins and progesterone
as prescribed in the PEPI Trial is accepted as the standard in the United States, the European Prospective Investigation results call for additional confirmation of endometrial protection with progestogens. It is essential to improve our understanding of the role of progestogens when added to estrogen therapy and the risk/benefit profile compared with estrogen alone in terms of the risks of diagnosis of endometrial and breast cancer. We hypothesize that estrogen acts via a mechanism that primes the tissue, and when progestogens are added, there is a promotional effect resulting in increased breast cancer diagnosis. The clinical characterization of progesterone and its differentiation from the progestins is an important component of the decision to add it to estrogen therapy.

Breast cancer is five times more common than endometrial cancer, and with regular monitoring, endometrial hyperplasia can be diagnosed well before endometrial cancer develops (313). In addition, the protective role of estrogen against risk of breast cancer, supported by the epidemiological literature, should not be overlooked in light of a potentially deleterious effect of progestogen.

Evaluation of thrombotic and thromboembolism risk, as well as the fact that different progestogens affect cardiovascular markers differently, suggests that progestosterone may have a decreased cardiovascular risk compared with other progestogens. The neuroprotective effect of progesterone is also being explored, whereas MPA has been shown to blunt estradiol’s beneficial effects in the brain.

A thorough review of the properties of the various progestogens shows that there are considerable data confirming the different effects of progestogens, the distinctiveness of progesterone, and the lack of a progestogen class effect. The properties of each progestogen should be carefully evaluated on an individual basis to determine its utility in postmenopausal HT.

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